

Volume 137, 2020, pp. 1–24 DOI: 10.1093/auk/ukaa030

RESEARCH ARTICLE

Sixty-first Supplement to the American Ornithological Society's *Check-list* of North American Birds

R. Terry Chesser,^{1,2,*} Shawn M. Billerman,³ Kevin J. Burns,⁴ Carla Cicero,⁵ Jon L. Dunn,⁶ Andrew W. Kratter,⁷ Irby J. Lovette,³ Nicholas A. Mason,⁸ Pamela C. Rasmussen,⁹ J. V. Remsen, Jr.,⁸ Douglas F. Stotz,¹⁰ and Kevin Winker¹¹

- ¹ U.S. Geological Survey, Patuxent Wildlife Research Center, Laurel, Maryland, USA
- ² National Museum of Natural History, Smithsonian Institution, Washington, DC, USA
- ³ Cornell Laboratory of Ornithology, Ithaca, New York, USA
- ⁴ Department of Biology, San Diego State University, San Diego, California, USA
- ⁵ Museum of Vertebrate Zoology, University of California, Berkeley, California, USA
- ⁶ 24 Idaho Street, Bishop, California, USA
- ⁷ Florida Museum of Natural History, University of Florida, Gainesville, Florida, USA
- 8 Museum of Natural Science and Department of Biological Sciences, Louisiana State University, Baton Rouge, Louisiana, USA
- ⁹ Michigan State University Museum and Department of Integrative Biology, East Lansing, Michigan, USA
- ¹⁰ Science & Education, Field Museum of Natural History, Chicago, Illinois, USA
- 11 University of Alaska Museum, Fairbanks, Alaska, USA
- * Corresponding author: chessert@si.edu

Authors are members of the Committee on Classification and Nomenclature—North and Middle America, of the American Ornithological Society (formerly American Ornithologists' Union), listed alphabetically after the Chairman.

Published June 30, 2020

This is the 20th supplement since publication of the 7th edition of the *Check-list of North American Birds* (American Ornithologists' Union [AOU] 1998). It summarizes decisions made between April 15, 2019 and April 15, 2020 by the American Ornithological Society's (formerly American Ornithologists' Union's) Committee on Classification and Nomenclature—North and Middle America. The Committee has continued to operate in the manner outlined in the 42nd Supplement (Banks et al. 2000). During the past year, Shawn M. Billerman and Nicholas A. Mason were added to the committee.

Changes in this supplement include the following: (1) 3 species (Buteo rufinus, Brotogeris chiriri, and Lanius collurio) are added to the main list, including 1 species transferred from the Appendix, on the basis of new distributional information; (2) 2 species (Anas diazi and Formicarius moniliger) are added to the main list because of splits from species already on the list; (3) 1 species (Zimmerius parvus) is added to the main list because of a split from a species already on the list, as well as from 2 extralimital species; (4) 2 scientific names are changed (to Sarkidiornis sylvicola and Turdus eunomus) because of splits from extralimital species, although the English names are retained; (5) the English name and distributional statement of 1 species (Zosterops japonicus) are changed because of a split from an extralimital species; (6) the distributional statement of 1 species (Thalasseus maximus)

is changed because of a split from an extralimital species; (7) 1 species (*Corvus caurinus*) is lost by merger with a species already on the list; (8) 1 species (Uraeginthus bengalus) is removed from the main list and placed in the Appendix; (9) 7 genera (Gymnasio, Poliocrania, Sipia, Dendroma, Pseudopipra, Helopsaltes, and Loriotus) are added due to splits from other genera, resulting in changes to 7 scientific names (Gymnasio nudipes, Poliocrania exsul, Sipia laemosticta, Dendroma rufa, Pseudopipra pipra, Helopsaltes ochotensis, and Loriotus luctuosus); (10) 1 genus (Atthis) is lost by merger with a genus already on the list, resulting in changes to 2 scientific names (Selasphorus heloisa and S. ellioti); (11) the gender ending of the scientific name of 1 species (Cyanolyca nanus) is corrected; (12) the English name of 1 species (*Epinecrophylla fulviventris*) is changed; (13) 1 new species (Alcedo atthis) is added to the Appendix; (14) 5 species (Anser anser, Coccyzus melacoryphus, Haematopus ostralegus, Pluvialis apricaria, and Pseudobulweria rostrata) are added to the list of species known to occur in the United States; and (15) 4 species (Numida meleagris, Estrilda melpoda, E. troglodytes, and Lonchura malacca) are removed from the list of species known to occur in the United States.

More sweeping changes derive from adoption of a new classification for a portion of the hummingbird subfamily Trochilinae, encompassing the genera *Chlorostilbon* through *Hylocharis*, which results in the following: (1) 7

species (Cynanthus auriceps, C. forficatus, C. canivetii, Microchera cupreiceps, M. chionura, Goldmania bella, and Eupherusa ridgwayi) are transferred to currently recognized genera; (2) 9 genera (Phaeoptila, Riccordia, Basilinna, Pampa, Leucolia, Saucerottia, Chrysuronia, Polyerata, and Chlorestes) are added because of splits from other genera; (3) 6 genera (Cyanophaia, Elvira, Goethalsia, Lepidopyga, Hylocharis, and Juliamyia) are deleted, 5 of which (all except Hylocharis) are subsumed into other genera; and (4) a new linear sequence is adopted for these genera and species.

Three subfamilies (Phasianinae, Tetraoninae, and Meleagridinae) are deleted, and new linear sequences are adopted for families in the order Suliformes, genera in the family Rallidae, species in the genera *Dendrortyx*, *Megascops* (and related genera), *Chloroceryle*, *Ara*, *Forpus*, *Myrmeciza* (and related genera), and *Progne*, and species in the families Phalacrocoracidae, Cathartidae, and Locustellidae, all due to new phylogenetic data.

Literature that provides the basis for the Committee's decisions is cited at the end of this supplement, and citations not already in the Literature Cited of the 7th edition (with supplements) become additions to it. A list of the bird species known from the AOS *Check-list* area can be found at http://checklist.americanornithology.org/taxa, and proposals that form the basis for this supplement can be found at https://americanornithology.org/nacc/current-prior-proposals/2020-proposals/.

The following changes to the 7th edition (page numbers refer thereto) and its supplements result from the Committee's actions:

p. xiii. New criteria are adopted for assessing the establishment of introduced species, largely following the revised standards of the American Birding Association (Pranty et al. 2008). In particular, the minimum persistence time for establishment has been changed from 10 to 15 yr, to allow for more accurate determination of the ability of an introduced species to persist in the wild.

Change the first sentence of the second paragraph under "Criteria for Inclusion" to the following: "Species that have been introduced by humans, either deliberately or accidentally, are considered to be established if the following criteria are met: (1) the species is documented by a specimen or published photograph, (2) there are persistent records for at least fifteen years, (3) the species is represented by a *bona fide* population rather than by scattered individuals, (4) the population can survive routine mortality and breeding failure, (5) the population is reasonably stable or increasing through successful reproduction, and (6) a publication documents the meeting of these criteria (*cf.* Roberson 1993, Pranty et al. 2008)."

p. xiii. Revised guidelines for English names are adopted, superseding those set forth in AOU (1983)

and modified in AOU (1998). Delete the current *English names* subsection in its entirety and replace it with the text available online at https://americanornithology.org/nacc/guidelines-for-english-bird-names/.

pp. xvii—liv. Increase the number in the title of the list of species to 2,158. Insert the following names in the proper position as indicated by the text of this supplement:

Sarkidiornis sylvicola Comb Duck.

Anas diazi Mexican Duck.

Selasphorus heloisa Bumblebee Hummingbird.

Selasphorus ellioti Wine-throated Hummingbird.

Phaeoptila sordida Dusky Hummingbird.

Riccordia ricordii Cuban Emerald.

†*Riccordia bracei* Brace's Emerald.

Riccordia swainsonii Hispaniolan Emerald.

Riccordia maugaeus Puerto Rican Emerald.

Riccordia bicolor Blue-headed Hummingbird.

Cynanthus auriceps Golden-crowned Emerald.

Cynanthus forficatus Cozumel Emerald.

Cynanthus canivetii Canivet's Emerald.

Basilinna leucotis White-eared Hummingbird.

Basilinna xantusii Xantus's Hummingbird.

Pampa curvipennis Wedge-tailed Sabrewing.

Dans a qualless I am tailed Cabraving

Pampa excellens Long-tailed Sabrewing.

Pampa rufa Rufous Sabrewing.

Microchera cupreiceps Coppery-headed Emerald.

Microchera chionura White-tailed Emerald.

Goldmania bella Pirre Hummingbird.

Eupherusa ridgwayi Mexican Woodnymph.

Leucolia violiceps Violet-crowned Hummingbird.

Leucolia viridifrons Green-fronted Hummingbird.

Saucerottia cyanocephala Azure-crowned Hummingbird.

Saucerottia hoffmanni Blue-vented Hummingbird.

Saucerottia beryllina Berylline Hummingbird.

Saucerottia cyanura Blue-tailed Hummingbird.

Saucerottia edward Snowy-bellied Hummingbird.

Chrysuronia coeruleogularis Sapphire-throated Hummingbird.

Chrysuronia humboldtii Humboldt's Sapphire.

Polyerata amabilis Blue-chested Hummingbird.

Polyerata decora Charming Hummingbird.

Chlorestes candida White-bellied Emerald.

Chlorestes eliciae Blue-throated Goldentail.

Chlorestes julie Violet-bellied Hummingbird.

Buteo rufinus Long-legged Buzzard. (A)

Gymnasio nudipes Puerto Rican Owl.

Brotogeris chiriri Yellow-chevroned Parakeet. (I)

Epinecrophylla fulviventris Checker-throated Stipplethroat.

Poliocrania exsul Chestnut-backed Antbird.

Sipia laemosticta Dull-mantled Antbird.

Formicarius moniliger Mayan Antthrush.

Dendroma rufa Buff-fronted Foliage-gleaner.

Pseudopipra pipra White-crowned Manakin.

Zimmerius vilissimus Guatemalan Tyrannulet.

Zimmerius parvus Mistletoe Tyrannulet.

Lanius collurio Red-backed Shrike. (A)

Cyanolyca nanus Dwarf Jay.

Zosterops japonicus Warbling White-eye. (H, I)

Helopsaltes ochotensis Middendorff's Grasshopper-

Warbler. (A)

Turdus eunomus Dusky Thrush. (A)

Loriotus luctuosus White-shouldered Tanager.

Delete the following names:

Sarkidiornis melanotos Comb Duck.

Phasianinae

Tetraoninae

Meleagridinae

Atthis heloisa Bumblebee Hummingbird.

Atthis ellioti Wine-throated Hummingbird.

Chlorostilbon auriceps Golden-crowned Emerald.

Chlorostilbon forficatus Cozumel Emerald.

Chlorostilbon canivetii Canivet's Emerald.

Chlorostilbon ricordii Cuban Emerald.

†Chlorostilbon bracei Brace's Emerald.

Chlorostilbon swainsonii Hispaniolan Emerald.

Chlorostilbon maugaeus Puerto Rican Emerald.

Cynanthus sordidus Dusky Hummingbird.

Cyanophaia bicolor Blue-headed Hummingbird.

Campylopterus curvipennis Wedge-tailed Sabrewing.

Campylopterus excellens Long-tailed Sabrewing.

Campylopterus rufus Rufous Sabrewing.

Elvira chionura White-tailed Emerald.

Elvira cupreiceps Coppery-headed Emerald.

Thalurania ridgwayi Mexican Woodnymph.

Amazilia candida White-bellied Emerald.

Amazilia amabilis Blue-chested Hummingbird.

Amazilia decora Charming Hummingbird.

Amazilia cyanocephala Azure-crowned Hummingbird.

Amazilia beryllina Berylline Hummingbird.

Amazilia cyanura Blue-tailed Hummingbird.

Amazilia hoffmanni Blue-vented Hummingbird.

Amazilia edward Snowy-bellied Hummingbird.

Amazilia violiceps Violet-crowned Hummingbird.

Amazilia viridifrons Green-fronted Hummingbird.

Goethalsia bella Pirre Hummingbird.

Lepidopyga coeruleogularis Sapphire-throated

Hummingbird.

Juliamyia julie Violet-bellied Hummingbird.

Hylocharis humboldtii Humboldt's Sapphire.

Hylocharis eliciae Blue-throated Goldentail.

Hylocharis leucotis White-eared Hummingbird.

Hylocharis xantusii Xantus's Hummingbird. *Megascops nudipes* Puerto Rican Screech-Owl.

Epinecrophylla fulviventris Checker-throated Antwren.

Myrmeciza exsul Chestnut-backed Antbird.

Myrmeciza laemosticta Dull-mantled Antbird.

Philydor rufum Buff-fronted Foliage-gleaner.

Dixiphia pipra White-crowned Manakin.

Zimmerius vilissimus Paltry Tyrannulet.

Cyanolyca nana Dwarf Jay.

Corvus caurinus Northwestern Crow.

Zosterops japonicus Japanese White-eye. (H, I)

Locustella ochotensis Middendorff's Grasshopper-

Warbler. (A)

Turdus naumanni Dusky Thrush. (A)

Uraeginthus bengalus Red-cheeked Cordonbleu. (H, I)

Tachyphonus luctuosus White-shouldered Tanager.

Adopt the following linear sequence for species in the genus *Dendrortyx*:

Dendrortyx leucophrys

Dendrortyx macroura

Dendrortyx barbatus

Adopt the following linear sequence for species in the family Phasianidae:

Meleagris gallopavo

Meleagris ocellata

Bonasa umbellus

Falcipennis canadensis

Lagopus lagopus

Lagopus muta

Lagopus leucura

Centrocercus urophasianus

Centrocercus minimus

Dendragapus obscurus

Dendragapus fuliginosus

Tympanuchus phasianellus

Tympanuchus cupido

Tympanuchus pallidicinctus

Perdix perdix

Phasianus colchicus

Lophura leucomelanos

Pavo cristatus

Francolinus pondicerianus

Francolinus francolinus

Gallus gallus

Tetraogallus himalayensis

Alectoris chukar

Coturnix japonica

Pternistis erckelii

Adopt the following linear sequence for species in the genus *Selasphorus*:

Selasphorus calliope Selasphorus rufus

Selasphorus sasin

Selasphorus platycercus

Selasphorus heloisa

Selasphorus ellioti

Selasphorus flammula

Selasphorus scintilla

Selasphorus ardens

Adopt the following linear sequence for species currently listed from *Chlorostilbon auriceps* through *Hylocharis xantusii* in the Trochilinae, adding asterisks before 2 species to indicate their uncertain generic placement:

Phaeoptila sordida Riccordia ricordii †Riccordia bracei Riccordia swainsonii Riccordia maugaeus Riccordia bicolor Cynanthus latirostris Cynanthus auriceps Cynanthus forficatus Cynanthus canivetii Chlorostilbon assimilis Basilinna leucotis Basilinna xantusii Pampa curvipennis Pampa excellens Pampa rufa Abeillia abeillei Klais guimeti

Orthorhyncus cristatus Campylopterus hemileucurus

Chalybura urochrysia Chalybura buffonii Thalurania colombica Microchera albocoronata Microchera cupreiceps Microchera chionura Goldmania violiceps Goldmania bella Eupherusa ridgwayi Eupherusa poliocerca Eupherusa cyanophrys Eupherusa eximia Eupherusa nigriventris Phaeochroa cuvierii Trochilus polytmus Leucolia violiceps Leucolia viridifrons Saucerottia cyanocephala Saucerottia hoffmanni Saucerottia beryllina

Saucerottia edward Amazilia rutila Amazilia yucatanensis Amazilia tzacatl *Amazilia luciae

*Amazilia boucardi

Saucerottia cyanura

Chrysuronia coeruleogularis Chrysuronia humboldtii Polyerata amabilis Polyerata decora Chlorestes candida Chlorestes eliciae Chlorestes julie

Adopt the following linear sequence for genera in the family Rallidae:

Neocrex Cyanolimnas Pardirallus Amaurolimnas Aramides Rallus Crex Porzana Gallinula Fulica Porphyrio Micropygia **Coturnicops Hapalocrex** Laterallus Zapornia

Adopt the following linear sequence for families in the order Suliformes:

FREGATIDAE SULIDAE ANHINGIDAE PHALACROCORACIDAE

Adopt the following linear sequence for species in the family Phalacrocoracidae:

Phalacrocorax penicillatus Phalacrocorax urile Phalacrocorax pelagicus Phalacrocorax carbo Phalacrocorax auritus Phalacrocorax brasilianus

Adopt the following linear sequence for species in the family Cathartidae:

Gymnogyps californianus Sarcoramphus papa Coragyps atratus Cathartes aura Cathartes burrovianus Adopt the following linear sequence for species in the genera *Psiloscops, Gymnasio*, and *Megascops*:

Psiloscops flammeolus
Gymnasio nudipes
Megascops trichopsis
Megascops clarkii
Megascops choliba
Megascops barbarus
Megascops cooperi
Megascops kennicottii
Megascops asio
Megascops seductus
Megascops guatemalae
Megascops centralis

Adopt the following linear sequence for species in the genus *Chloroceryle*:

Chloroceryle amazona Chloroceryle aenea Chloroceryle americana Chloroceryle inda

Adopt the following linear sequence for species in the genus *Ara*:

Ara ararauna Ara severus Ara tricolor Ara macao Ara chloropterus Ara militaris Ara ambiguus

Adopt the following linear sequence for species in the genus *Forpus*:

Forpus cyanopygius Forpus passerinus Forpus conspicillatus

Adopt the following linear sequence for species currently or formerly in *Myrmeciza*, and insert an asterisk to indicate the uncertain generic placement of *Myrmeciza zeledoni*:

Myrmeciza longipes *Myrmeciza zeledoni Poliocrania exsul Sipia laemosticta

Adopt the following linear sequence for species in the genus *Progne*:

Progne tapera Progne subis Progne elegans Progne chalybea Progne sinaloae

Progne cryptoleuca Progne dominicensis

Adopt the following linear sequence for species in the family Locustellidae:

Helopsaltes ochotensis Locustella lanceolata Locustella fluviatilis

Note: The entries below follow the current linear sequence as established in this and previous supplements, although entries continue to be cross-referenced to page numbers in AOU (1998).

1. [p. 58] A record of *Anser anser* in the United States is recognized. Change the last paragraph of the distributional statement to the following:

Accidental in Greenland, Newfoundland, Nova Scotia, and Quebec. Accidental in Connecticut (Wallingford, New Haven County, 22 February–8 March 2009; photos; Kaplan and Hanisek 2012).

2. [p. 58] *Sarkidiornis sylvicola* is treated as a species separate from extralimital species *S. melanotos*. Remove the species account for *S. melanotos* and replace it with the following new account:

Sarkidiornis sylvicola Ihering and Ihering. Comb Duck.

Sarkidiornis sylvicola Ihering and Ihering, 1907, in Museu Paulista, São Paulo, Catálogo Fauna Brasileira
1: 72. (Iguapé, São Paulo, Brazil, and Buenos Aires, Argentina.) New name for Anas carunculata
Lichtenstein, 1819, preoccupied by Anas carunculata
Vieillot, 1816, Nouvelle Dictionnaire Histoire
Naturelle, nouv. éd., vol. 5, p. 109.

Habitat.—Freshwater Lakes and Ponds, Freshwater Marshes (0–1200 m; Tropical to Temperate zones).

Distribution.—Resident locally in tropical America from eastern Panama (Río Chucunaque in eastern Darién, casually west to La Jagua, eastern Panamá province), south through northern South America to northwestern Peru, southeastern Bolivia, northern Argentina, and Uruguay (generally absent from Amazonia).

Notes.—Formerly (e.g., AOU 1983, 1998) considered conspecific with *S. melanotos* (Pennant, 1769) [Knob-billed Duck], but separated based on differences in plumage and because the original lump of these species (by Delacour and Mayr 1945) was based on hybridization in captivity.

3. [p. 68] *Anas diazi* is treated as a species separate from *A. platyrhynchos*. Change the species account for *A. platyrhynchos* as follows: delete mention of the *diazi* group from the habitat and distributional statements and change the Notes to: "The *Anas platyrhynchos* complex

includes 14 closely related species; A. platyrhynchos appears to be most closely related to the New World radiation, which includes A. diazi, A. fulvigula, A. rubripes, A. wyvilliana, and A. laysanensis, and to A. poecilorhyncha J. R. Forster 1781 [Indian Spot-billed Duck] and A. zonorhyncha Swinhoe, 1866 [Eastern Spot-billed Duck] in the Old World (Lavretsky et al. 2014a). In various older treatments, some or even all New World taxa were treated as conspecific under the name A. platyrhynchos (e.g., Johnsgard 1961, 1967). Anas rubripes and A. fulvigula hybridize frequently with A. platyrhynchos in an area of broad overlap, largely as a result of introductions and range expansions of the latter into the range of A. rubripes and A. fulvigula. These 3 forms differ somewhat behaviorally and tend to segregate as species (Brodsky and Weatherhead 1984, Brodsky et al. 1988, Hepp et al. 1988, Ford et al. 2017, Lavretsky et al. 2019b), but early genetic studies found them difficult to differentiate (Ankney et al. 1986, Ankney and Dennis 1988, Avise et al. 1990, McCracken et al. 2001, Lavretsky et al. 2014a, b). More recent genomic studies have found that they are genetically separable (Lavretsky et al. 2015, 2019a,b), with differences likely the result of selection and demographic processes (Kirby et al. 2004, Lavretsky et al. 2019b). Further, genetic evidence suggests that hybridization is not as widespread as previously believed (Ford et al. 2017), and that A. platyrhynchos and A. rubripes do not represent a hybrid swarm (Lavretsky et al. 2019b). See comments under A. diazi."

Insert the following new species account after the account for *A. platyrhynchos*:

Anas diazi Ridgway. Mexican Duck.

Anas diazi Ridgway, 1886, Auk 3: 332. (San Ysidro, Puebla, Mexico.)

Habitat.—Freshwater Marshes (0–2500 m).

Distribution.—*Breeds* from southeastern Arizona, southern New Mexico, and west-central Texas south in the highlands of Mexico to Jalisco, Michoacán, México, Distrito Federal, Tlaxcala, and Puebla.

Winters in the breeding range and east to southern Coahuila, San Luis Potosí, and eastern Tamaulipas.

Nonbreeding birds occur casually throughout the year north through much of Colorado and in Utah north to Great Salt Lake, west to the Lower Colorado River Valley, and east to the Lower Rio Grande Valley. Accidental west to San Luis Obispo County, California, north to Albany County, Wyoming, and east to southwestern Nebraska. Difficulties distinguishing this species from *A. fulvigula* may be decreasing detection east of its usual range.

Notes.—Formerly (e.g., AOU 1983, 1998) considered conspecific with *A. platyrhynchos*, although prior to this (until AOU 1973) the 2 were treated as separate species. Newly separated based on assortative mating in the narrow contact zone between these species (Bellrose 1976, Hubbard

1977, Brown 1985) and genomic data that indicate restricted gene flow between them (Lavretsky et al. 2015, 2019a).

4. [p. 123] Records of *Numida meleagris* in the United States are recognized as belonging to populations that were never established (Pratt et al. 1987; *contra* Walker 1967, Berger 1981, AOU 1983, 1998). Remove this species from the list of species known to occur in the United States, remove "in the Hawaiian Islands (in 1874 on Hawaii and possibly other main islands, perhaps not well established)," from the second paragraph of the distributional statement, and add the following paragraph to the end of the distributional statement:

Introduced in the Hawaiian Islands (in 1874 on Kauai; many later introductions on Kauai and other main islands), but populations failed to become established (Pyle and Pyle 2017).

5. [pp. 123–124] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Tsai et al. 2019) have shown that our current linear sequence of species in the genus *Dendrortyx* does not reflect their evolutionary relationships. These findings result in the following changes:

After the heading and citation for *Dendrortyx*, insert the following:

Notes.—Linear sequence of species follows Tsai et al. (2019).

Rearrange the sequence of species in *Dendrortyx* to:

Dendrortyx leucophrys Dendrortyx macroura Dendrortyx barbatus

6. [pp. 114–123] Phylogenetic analyses of nuclear and mitochondrial DNA sequences and morphological data (Crowe et al. 2006, Wang et al. 2013, Hosner et al. 2016) have shown that our current subfamily structure and linear sequence of species in the family Phasianidae do not reflect their evolutionary relationships. These findings result in the following changes:

Delete the heading Subfamily PHASIANINAE: Partridges and Pheasants, and the headings and Notes for Subfamily TETRAONINAE: Grouse, and Subfamily MELEAGRIDINAE: Turkeys. Insert the following Notes under the heading and citation for Phasianidae:

Notes.—Formerly divided into subfamilies Phasianinae, Tetraoninae, and Meleagridinae, but analyses of genetic and morphological data (Crowe et al. 2006, Wang et al. 2013, Hosner et al. 2016) indicate that the species formerly included in Tetraoninae and Meleagridinae are embedded within the Phasianinae.

Rearrange the sequence of species in the Phasianidae to:

Meleagris gallopavo Meleagris ocellata Bonasa umbellus Falcipennis canadensis Lagopus lagopus Lagopus muta Lagopus leucura Centrocercus urophasianus Centrocercus minimus Dendragapus obscurus Dendragapus fuliginosus Tympanuchus phasianellus Tympanuchus cupido Tympanuchus pallidicinctus Perdix perdix Phasianus colchicus Lophura leucomelanos Pavo cristatus Francolinus pondicerianus Francolinus francolinus Gallus gallus Tetraogallus himalayensis Alectoris chukar Coturnix japonica Pternistis erckelii

7. [p. 248] Records of *Coccyzus melacoryphus* in the United States are recognized. Delete the last sentence of the distributional statement and add the following new paragraph:

Accidental in southern Texas (brought to a rehabilitation center in Weslac, Hidalgo County, 10 February 1986; specimen, LSUMNS; Lockwood 1999, Pyle et al. 2019) and in southern Florida (Delray Beach, Palm Beach County, 6–10 February 2019; photos; Pyle et al. 2019, Kratter et al. 2020).

8. [pp. 311–312] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (McGuire et al. 2014, Licona-Vera and Ornelas 2017) have shown that *Selasphorus* is paraphyletic with respect to *Atthis*, and that our current linear sequence of species in the genus *Selasphorus* does not reflect their evolutionary relationships. These findings result in the following changes:

Delete the heading Genus *ATTHIS* Reichenbach and the Notes under this heading, move the citation for *Atthis* into the synonymy of *Selasphorus*, and change the Notes under *Selasphorus* to the following:

Notes.—See comments under *S. heloisa*. Sequence of species follows McGuire et al. (2014) and Licona-Vera and Ornelas (2017).

Change *Atthis heloisa* to *Selasphorus heloisa* and *Atthis ellioti* to *Selasphorus ellioti*, add parentheses around the authority for *S. ellioti*, make the appropriate change in the generic abbreviation within the existing Notes for *S. ellioti*, and change the Notes under *S. heloisa* to the following:

Notes.—Also known as Heloise's Hummingbird. Formerly (AOU 1983, 1998) placed in *Atthis* with sister species *S. ellioti*, but genetic data (McGuire et al. 2014, Licona-Vera and Ornelas 2017) indicate that *Selasphorus* as previously constituted was paraphyletic with respect to *Atthis*, as anticipated by Howell and Webb (1995).

Rearrange the sequence of species in Selasphorus to:

Selasphorus calliope Selasphorus rufus Selasphorus sasin Selasphorus platycercus Selasphorus heloisa Selasphorus ellioti Selasphorus flammula Selasphorus scintilla Selasphorus ardens

9. [pp. 289–303] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (McGuire et al. 2014, Hernández-Baños et al. 2020) have shown that the generic limits and linear sequence of species in a portion of the subfamily Trochilinae (genera from *Chlorostilbon* through *Hylocharis*) do not accurately reflect their evolutionary relationships. We adopt a new classification based on their conclusions, which results in the following changes:

Rearrange the sequence of genera and species currently listed from *Chlorostilbon auriceps* through *Hylocharis xantusii* as follows, in keeping with the genus transfers detailed below and adding parentheses to the author names for *Cynanthus forficatus*, *Pampa rufa*, *Eupherusa ridgwayi*, and *Saucerottia cyanura*, and removing parentheses from the author names for *Pampa excellens* and *Polyerata decora*:

Genus Phaeoptila Gould

Phaeoptila sordida Gould

Genus Riccordia Reichenbach

Riccordia ricordii (Gervais)

Riccordia bracei (Lawrence)

Riccordia swainsonii (Lesson)

Riccordia maugaeus (Audebert and Vieillot)

Riccordia bicolor (Gmelin)

Genus Cynanthus Swainson

Cynanthus latirostris Swainson

Cynanthus auriceps (Gould)

Cynanthus forficatus (Ridgway)

Cynanthus canivetii (Lesson)

Genus Chlorostilbon Gould

Chlorostilbon assimilis Lawrence

Genus Basilinna Boie

Basilinna leucotis (Vieillot)

Basilinna xantusii (Lawrence)

Genus Pampa Reichenbach

Pampa curvipennis (Deppe)

Pampa excellens Wetmore

Pampa rufa (Lesson)

Genus Abeillia Bonaparte

Abeillia abeillei (DeLattre and Lesson)

Genus Klais Reichenbach

Klais guimeti (Bourcier)

Genus Orthorhyncus Lacépède

Orthorhyncus cristatus (Linnaeus)

Genus Campylopterus Swainson

Campylopterus hemileucurus (Deppe)

Genus Chalybura Reichenbach

Chalybura urochrysia (Gould)

Chalybura buffonii (Lesson)

Genus Thalurania Gould

Thalurania colombica (Bourcier)

Genus Microchera Gould

Microchera albocoronata (Lawrence)

Microchera cupreiceps (Lawrence)

Microchera chionura (Gould)

Genus Goldmania Nelson

Goldmania violiceps Nelson

Goldmania bella (Nelson)

Gotamania betta (Neisott)

Genus Eupherusa Gould

Eupherusa ridgwayi (Nelson)

Eupherusa poliocerca Elliot

Eupherusa cyanophrys Rowley and Orr

Eupherusa eximia (DeLattre)

Eupherusa nigriventris Lawrence

Genus Phaeochroa Gould

Phaeochroa cuvierii (DeLattre and Bourcier)

Genus Trochilus Linnaeus

Trochilus polytmus Linnaeus

Genus *Leucolia* Mulsant, Verreaux and Verreaux

Leucolia violiceps (Gould)

Leucolia viridifrons (Elliot)

Genus Saucerottia Bonaparte

Saucerottia cyanocephala (Lesson)

Saucerottia hoffmanni (Cabanis and Heine)

Saucerottia beryllina (Deppe)

Saucerottia cyanura (Gould)

Saucerottia edward (DeLattre and Bourcier)

Genus Amazilia Lesson

Amazilia rutila (DeLattre)

Amazilia yucatanensis (Cabot)

Amazilia tzacatl (De la Llave)

Amazilia luciae (Lawrence)

Amazilia boucardi (Mulsant)

Genus Chrysuronia Bonaparte

Chrysuronia coeruleogularis (Gould)

Chrysuronia humboldtii (Bourcier and Mulsant)

Genus Polyerata Heine

Polyerata amabilis (Gould)

Polyerata decora Salvin

Genus Chlorestes Reichenbach

Chlorestes candida (Bourcier and Mulsant)

Chlorestes eliciae (Bourcier and Mulsant)

Chlorestes julie (Bourcier)

Remove the citation for *Phaeoptila* from the synonymy of *Cynanthus*, and insert the following new heading, citation, and Notes after the species account for *Selasphorus ardens*:

Genus PHAEOPTILA Gould

Phaeoptila Gould, 1861, A Monograph of the Trochilidae, part 5, text to plate 340. Type, by original designation, *Cyanomyia sordida* Gould.

Notes.—Formerly (e.g., AOU 1983, 1998) included in *Cynanthus*, but see Stiles et al. (2017) for resurrection of *Phaeoptila* based on genetic data (McGuire et al. 2014).

Change *Cynanthus sordidus* to *Phaeoptila sordida*, move the account for this species to follow the heading and citation for *Phaeoptila*, and insert the following Notes at the end of the species account:

Notes.—See comments under Phaeoptila.

Insert the following new heading, citation, and Notes after the species account for *Phaeoptila sordida*:

Genus RICCORDIA Reichenbach

Chlorestes δ Riccordia Reichenbach, 1854, Journal für Ornithologie 1 (Beiliegend zu Extraheft): 8. Type, by subsequent designation (G. R. Gray, 1855), Riccordia ramondii Reichenbach = Ornismya ricordii Gervais.

Notes.—Formerly (e.g., AOU 1983, 1998) included in *Chlorostilbon*, but see Stiles et al. (2017) for resurrection of *Riccordia* based on genetic data (McGuire et al. 2014).

Change the generic names of *Chlorostilbon ricordii*, *C. bracei*, *C. swainsonii*, *C. maugaeus*, and *Cyanophaia bicolor* to *Riccordia*; delete the genus heading for *Cyanophaia*; move the citation for *Cyanophaia* into the synonymy of *Riccordia*; make the appropriate changes in generic names or abbreviations within the existing Notes; and place the accounts for these species under the heading and citation for *Riccordia*.

Insert the following Notes under the heading Genus *CHLOROSTILBON* Gould:

Notes.—See comments under Riccordia and Cynanthus.

Transfer *Chlorostilbon auriceps*, *Chlorostilbon forficatus*, and *Chlorostilbon canivetii* to the genus *Cynanthus*, make

the appropriate changes to generic names within the existing Notes, and replace the last two sentences of the existing Notes for *C. canivetti* with the following: Formerly (e.g., AOU 1998), along with *C. auriceps* and *C. forficatus*, placed in *Chlorostilbon*, but see Stiles et al. (2017) for transfer of these species to *Cynanthus* based on genetic data (McGuire et al. 2014).

Remove the citation for *Basilinna* from the synonymy of *Hylocharis*, and insert the following new heading, citation, and Notes after the species account for *Chlorostilbon assimilis*:

Genus BASILINNA Boie

Basilinna Boie, 1831, Isis von Oken 1831: col. 546. Type, by subsequent designation (G. R. Gray, 1855), *Trochilus leucotis* Vieillot.

Notes.—Formerly (e.g., AOU 1983, 1998) included in *Hylocharis*, but see Stiles et al. (2017) for resurrection of *Basilinna* based on genetic data (McGuire et al. 2014), as anticipated by Howell and Webb (1995).

Change *Hylocharis leucotis* to *Basilinna leucotis* and *Hylocharis xantusii* to *Basilinna xantusii*, move the accounts for these species to follow the heading and citation for *Basilinna*, and replace the existing Notes for both species with the following:

Notes.—See comments under Basilinna.

Remove the citation for *Pampa* from the synonymy of *Campylopterus*, and insert the following new heading, citation, and Notes after the species account for *Basilinna xantusii*:

Genus PAMPA Reichenbach

Pampa Reichenbach, 1854, Journal für Ornithologie
1 (Beiliegend zu Extraheft): 11. Type, by monotypy,
P. campyloptera Reichenbach = Ornismya pampa
Lesson = Trochilus curvipennis Deppe.

Notes.—Formerly (e.g., AOU 1983, 1998) included in *Campylopterus*, but see Stiles et al. (2017) for resurrection of *Pampa* based on genetic data (McGuire et al. 2014).

Change Campylopterus curvipennis to Pampa curvipennis, Campylopterus excellens to Pampa excellens, and Campylopterus rufus to Pampa rufa; place the accounts for these species under the heading and citation for Pampa; make the appropriate changes in generic names or abbreviations within the existing Notes; and either insert the following Notes (for P. rufa) or add the following

sentence to the end of the existing Notes (for *P. curvipennis* and *P. excellens*): See comments under *Pampa*.

Insert the following Notes under the heading Genus *CAMPYLOPTERUS* Swainson:

Notes.—See comments under Pampa.

Transfer *Elvira chionura* and *E. cupreiceps* to the genus *Microchera*, delete the genus heading for *Elvira*, move the citation for *Elvira* into the synonymy of *Microchera*, and insert the following Notes at the end of the species accounts for *M. chionura* and *M. cupreiceps*:

Notes.—Formerly (e.g., AOU 1983, 1998) placed in *Elvira*, but see Stiles et al. (2017) for transfer of these species to *Microchera* based on genetic data (McGuire et al. 2014).

Transfer *Goethalsia bella* to the genus *Goldmania*, delete the genus heading for *Goethalsia*, move the citation for *Goethalsia* into the synonymy of *Goldmania*, and insert the following sentence at the beginning of the existing Notes for *G. bella*: Formerly (e.g., AOU 1983, 1998) placed in *Goethalsia*, but see Stiles et al. (2017) for transfer of this species to *Goldmania* based on genetic data (McGuire et al. 2014).

Transfer *Thalurania ridgwayi* to the genus *Eupherusa*, and insert the following sentence at the end of the existing Notes for this species: Formerly (e.g., AOU 1998) placed in *Thalurania*, but see Stiles et al. (2017) for transfer of this species to *Eupherusa* based on genetic data (McGuire et al. 2014).

Insert the following new heading, citation, and Notes after the species account for *Trochilus polytmus*:

Genus LEUCOLIA Mulsant, Verreaux and Verreaux

Leucolia Mulsant, and J. and E. Verreaux, 1866, Mémoires Société Impériale Sciences Naturelles de Cherbourg 12: 174. Type, by subsequent designation (Elliot, 1897; Stiles et al., 2017), Cyanomyia viridifrons Elliot.

Notes.—Formerly (e.g., AOU 1983, 1998) included in *Amazilia*, but see Stiles et al. (2017) for resurrection of *Leucolia* based on genetic data (McGuire et al. 2014).

Change *Amazilia violiceps* to *Leucolia violiceps* and *Amazilia viridifrons* to *Leucolia viridifrons*, move the accounts for these species to follow the heading and citation for *Leucolia*, make the appropriate changes in generic names or abbreviations within the existing Notes, and insert the following at the end of the existing Notes for each species: See comments under *Leucolia*.

Remove the citation for *Saucerottia* from the synonymy of *Amazilia*, and insert the following new heading, citation, and Notes after the species account for *Leucolia viridifrons*:

Genus SAUCEROTTIA Bonaparte

Saucerottia Bonaparte, 1850, Conspectus Generum Avium 1(1): 77. Type, by original designation, Saucerottia typica Bonaparte = Trochilus saucerrottei [sic] DeLattre and Bourcier.

Notes.—Formerly (e.g., AOU 1983, 1998) included in *Amazilia*, but see Stiles et al. (2017) for resurrection of *Saucerottia* based on genetic data (McGuire et al. 2014).

Change the generic names of *Amazilia cyanocephala*, *A. hoffmanni*, *A. beryllina*, *A. cyanura*, and *A. edward* to *Saucerottia*; place the accounts for these species under the heading and citation for *Saucerottia*; make the appropriate changes in generic names or abbreviations within the existing Notes; and insert the following at the end of the existing Notes for each species: See comments under *Saucerottia*.

Insert the following Notes under the heading Genus *AMAZILIA* Lesson:

Notes.—See comments under *Leucolia*, *Saucerottia*, and *Polyerata*.

Insert the following at the end of the species account for *Amazilia luciae*:

Notes.—This species and *A. boucardi* are almost certainly unrelated to true *Amazilia*, but were not included in McGuire et al. (2014) and are of uncertain generic placement. These species are retained in *Amazilia* until they can be placed confidently based on new data.

Insert the following at the end of the existing Notes for *Amazilia boucardi*: See comments under *Amazilia luciae*.

Insert the following new heading, citation, and Notes after the species account for *Amazilia boucardi*:

Genus CHRYSURONIA Bonaparte

Chrysuronia Bonaparte, 1850, Conspectus Generum Avium 1: 75. Type, by subsequent designation (G. R. Gray, 1855), Ornismya oenone Lesson.

Notes.—Formerly considered an extralimital monotypic genus, but see Stiles et al. (2017) for transfer of *Lepidopyga*

coeruleogularis and Hylocharis humboldtii to Chrysuronia based on genetic data (McGuire et al. 2014).

Delete the genus heading for Lepidopyga and move the citation for Lepidopyga into the synonymy of Chrysuronia. Change Lepidopyga coeruleogularis to Chrysuronia coeruleogularis and Hylocharis humboldtii to Chrysuronia humboldtii, place the accounts for these species under the heading and citation for Chrysuronia, make the appropriate changes in generic names or abbreviations within the existing Notes, and either insert the following Notes (for C. coeruleogularis) or add the following sentence to the end of the existing Notes (for C. humboldtii): See comments under Chrysuronia.

Remove the citation for *Polyerata* from the synonymy of *Amazilia*, and insert the following new heading, citation, and Notes after the species account for *Chrysuronia humboldtii*:

Genus POLYERATA Heine

Polyerata Heine, 1863, Journal für Ornithologie 11: 194. Type, by monotypy, *Trochilus amabilis* Gould.

Notes.—Formerly (e.g., AOU 1983, 1998) included in *Amazilia*, but see Stiles et al. (2017) for resurrection of *Polyerata* based on genetic data (McGuire et al. 2014).

Change *Amazilia amabilis* to *Polyerata amabilis* and *Amazilia decora* to *Polyerata decora*, move the accounts for these species to follow the heading and citation for *Polyerata*, make the appropriate changes in generic names or abbreviations within the existing Notes, and insert the following at the end of the existing Notes: See comments under *Polyerata*.

Delete the genus heading and citation for *Hylocharis* and the genus heading for *Juliamyia*; move the citations for *Damophila*, *Juliamyia*, and *Neodamophila* into the synonymy of *Chlorestes*; and insert the following heading, citation, and Notes after the species account for *Polyerata decora*:

Genus CHLORESTES Reichenbach

Chlorestes Reichenbach, 1854, Journal für Ornithologie 1 (Beiliegend zu Extraheft): 7. Type, by subsequent designation (Salvin, 1892), Trochilus notatus Reich.

Notes.—Formerly considered an extralimital monotypic genus, but see Stiles et al. (2017) for transfer of *Amazilia candida*, *Hylocharis eliciae*, and *Juliamyia*

julie to *Chlorestes* based on genetic data (McGuire et al. 2014).

Change *Amazilia candida* to *Chlorestes candida*, *Hylocharis eliciae* to *Chlorestes eliciae*, and *Juliamyia julie* to *Chlorestes julie*; move the accounts for these species to follow the heading and citation for *Chlorestes*; and insert the following at the end of the species accounts for *C. candida* and *C. eliciae*:

Notes.—See comments under Chlorestes.

Change the Notes under Chlorestes julie to:

Notes.—Previously (e.g., AOU 1983, 1998) placed in *Damophila* Reichenbach, 1854, but this name is preoccupied by *Damophila* Curtis, 1832, a genus of Lepidoptera (Özdikmen 2008). Later (Chesser et al. 2017) transferred to *Juliamyia* Bonaparte, 1854, but genetic evidence indicates that it should be placed in *Chlorestes* (McGuire et al. 2014). See comments under *Chlorestes*.

10. [129–138] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Garcia-R et al. 2014, 2020) have shown that the current linear sequence of genera in the Rallidae does not reflect their evolutionary relationships.

After the heading Family **RALLIDAE**: Rails, Gallinules, and Coots, replace the existing Notes with the following:

Notes.—Linear sequence of genera follows Garcia-R et al. (2014, 2020).

Rearrange the sequence of genera in the Rallidae to:

Neocrex
Cyanolimnas
Pardirallus
Amaurolimnas
Aramides
Rallus
Crex
Porzana
Gallinula
Fulica
Porphyrio
Micropygia
Coturnicops

Hapalocrex

Laterallus

Zapornia

11. [p. 149] A record of *Haematopus ostralegus* in the United States is recognized. Substitute the following 2 paragraphs for the current final paragraph of the distributional statement:

Casual in Newfoundland (Fox Island near Tors Cove, Avalon Peninsula, 24–25 May 1994; photos; Mactavish

1994; Lushes Bight, Long Island, 15–23 May 2019; photos; e.g., https://ebird.org/checklist/S56448089; Eilliton, 5–9 April 2020; photos; e.g., https://ebird.org/checklist/S66745673). Sight report from Eastport.

Accidental in Alaska (Buldir Island, western Aleutians, 26 May–13 June 2012; photos; Gibson et al. 2013).

12. [p. 143] Records of *Pluvialis apricaria* in the United States are recognized. Change the final paragraph of the distributional statement to:

Casual in eastern North America in Labrador and Newfoundland, Quebec, Nova Scotia, Maine, New Jersey, and Delaware; and in Alaska (Ketchikan Airport, Ketchikan, 13–14 January 2001; specimen, UAM; Piston and Heinl 2001; St. Paul Island, Pribilof Islands, 24 January 2015; photos; https://ebird.org/checklist/S24051721; Barrow, 19 June 2017; photos; https://ebird.org/checklist/S37915248).

13. [pp. 197–198] *Thalasseus albididorsalis* is considered a species separate from *T. maximus*, resulting in the following changes to the species account for *T. maximus*:

In the breeding paragraph of the distributional statement change "in South America on the coast of northern Argentina; and in West Africa (islands off Mauritania)." to "and in South America on the coast of northern Argentina." Change the wintering paragraph of the distributional statement to "Winters from central California, the Gulf coast and North Carolina south along both coasts of the Americas to Peru, Uruguay, and Argentina." In the final paragraph of the distributional statement, change "also in the British Isles, Norway, Spain, Gibraltar, and Mozambique; a sight report from interior Mexico (Distrito Federal)." to "also in the British Isles, France, and Spain; a sight report from interior Mexico (Distrito Federal). Many European records of this species, including one from Norway, were identified as Thalasseus maximus sensu lato and their current species identification is unclear; however, most records from Gibraltar and Spain are believed to be of *T. albididorsalis* (Dufour and Crochet 2020)."

Replace the existing Notes with the following: Formerly (e.g., AOU 1983, 1998) considered conspecific with *T. albididorsalis* (Hartert, 1921) [West African Crested Tern], but separated based on genetic data that indicate that *T. albididorsalis* is sister to *T. bengalensis* (Lesson, 1831) [Lesser Crested Tern] (Collinson et al. 2017), and differences in vocalizations and morphology (summarized in Dufour and Crochet 2020).

14. [p. 16] Records of *Pseudobulweria rostrata* in the United States are recognized. Replace the last paragraph of the distributional statement with the following 2 paragraphs:

Rare in the eastern Pacific Ocean off Panama (southeast to the Azuero Peninsula), Costa Rica, El Salvador,

Guatemala, and western Mexico (north to the southern Gulf of California off Baja California Sur and Sinaloa).

Accidental off Hawaii (2 km west of Nā-wiliwili Harbor, Kauai; photos and measurements of bird in hand; VanderWerf et al. 2018) and off North Carolina (Hatteras, 29 May 2018; photos; https://ebird.org/checklist/S46146022).

15. [pp. 28–36] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (e.g., Ericson et al. 2006, Hackett et al. 2008, Prum et al. 2015) have shown that the current linear sequence of families in the Suliformes does not reflect their evolutionary relationships.

Insert the following at the end of the existing Notes under Order **SULIFORMES**: Frigatebirds, Boobies, Cormorants, Darters, and Allies: Linear sequence of families follows Ericson et al. (2006), Hackett et al. (2008), and Prum et al. (2015).

Rearrange the sequence of families in the Suliformes to:

FREGATIDAE SULIDAE ANHINGIDAE PHALACROCORACIDAE

16. [pp. 32–34] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Kennedy and Spencer 2014) have shown that our current linear sequence of species in the family Phalacrocoracidae does not reflect their evolutionary relationships. These findings result in the following changes:

After the heading Family **PHALACROCORACIDAE**: Cormorants, replace the existing Notes with the following: **Notes.**—Linear sequence of species follows Kennedy and Spencer (2014).

Rearrange the sequence of species in the family Phalacrocoracidae to:

Phalacrocorax penicillatus Phalacrocorax urile Phalacrocorax pelagicus Phalacrocorax carbo Phalacrocorax auritus Phalacrocorax brasilianus

17. [pp. 51–53] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Johnson et al. 2016) have shown that our current linear sequence of species in the family Cathartidae does not reflect their evolutionary relationships. These findings result in the following changes:

Insert the following Notes after the heading Family **CATHARTIDAE**: New World Vultures:

Notes.—Linear sequence of species follows Johnson et al. (2016).

Rearrange the sequence of species in the family Cathartidae to:

Gymnogyps californianus Sarcoramphus papa Coragyps atratus Cathartes aura Cathartes hurrovianus

18. [p. 102] After the species account for *Buteo regalis*, insert the following new species account:

Buteo rufinus (Cretzschmar). Long-legged Buzzard.

Falco rufinus Cretzschmar, 1827, in Rüppell, Atlas Reise Nördlichen Afrika, Vögel (1826), p. 40, pl. 27. (Upper Nubia, Shendi, Sennar, and Ethiopia.)

Habitat.—Primarily open, arid, semi-desert, uncultivated country with gorges and crags for nesting; also locally in woodlands. In winter open areas, including grasslands.

Distribution.—*Breeds* from Hungary, southern Ukraine, and Kazakhstan to northwestern China and northwestern Mongolia and south to the Balkans, Cyprus, Turkey, Sinai, Arabian Peninsula, Iraq, south-central Iran, western Afghanistan, northern Pakistan, and Kashmir.

Winters in southern part of breeding range and south in Nile Valley to Sudan, throughout Pakistan, northwestern India, and southern Tibet; rarely to Bhutan, central India, and Bangladesh.

Resident in northwestern Africa east to northwestern Libya and south to Mauritania.

Casual or accidental to North Sea coast, Finland, Nigeria, Senegal, Zanzibar, Sri Lanka, Myanmar, and Malaysia. Unverified reports from the Andaman Islands.

Accidental in Alaska (St. Paul Island, Pribilof Islands, 15 November 2018–7 April 2019; photos; Pyle et al. 2019).

19. [p. 254–257] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Dantas et al. 2016, Salter et al. 2020) have shown that *Megascops* is paraphyletic with respect to *Psiloscops*, and that our current linear sequence of species in these genera does not reflect their evolutionary relationships. These findings result in the following changes:

Remove the citation for *Gymnasio* from the synonymy of *Megascops*, and insert the following new heading, citation, and Notes after the species account for *Psiloscops flammeolus*:

Genus GYMNASIO Bonaparte

Gymnasio Bonaparte, 1854, Revue et Magasin de Zoologie (2)6: 543. Type, by monotypy [Strix] nudipes Daudin.

Notes.—See comments under *Gymnasio nudipes* and *Megascops*.

Change *Megascops nudipes* to *Gymnasio nudipes*, move the species account to follow the heading and citation for *Gymnasio*, change the English name to Puerto Rican Owl, and change the Notes under *G. nudipes* to the following:

Notes.—Formerly (e.g., Banks et al. 2003) placed in *Megascops*, but genetic data (Dantas et al. 2016, Salter et al. 2020) indicate that *Megascops* as previously constituted was paraphyletic and that *Gymnasio nudipes* is not part of *Megascops sensu stricto*. This species was also previously placed in *Otus* (e.g., AOU 1983, 1998). Also known as Puerto Rican Screech-Owl and Puerto Rican Bare-legged Owl. See comments under *Megascops*.

Insert the following sentence at the end of the existing Notes for *Megascops*: Linear sequence of species in *Psiloscops*, *Gymnasio*, and *Megascops* follows Dantas et al. (2016).

Replace the existing Notes under Genus *PSILOSCOPS* Coues with the following:

Notes.—Formerly (e.g., AOU 1983, 1998) merged with *Otus* but now treated as separate based on genetic data that show it, along with *Gymnasio*, to be sister to *Megascops* (Proudfoot et al. 2007, Wink et al. 2009, Dantas et al. 2016). See comments under *Megascops*.

Rearrange the sequence of species in *Psiloscops*, *Gymnasio*, and *Megascops* to:

Psiloscops flammeolus Gymnasio nudipes Megascops trichopsis Megascops clarkii Megascops choliba Megascops barbarus Megascops cooperi Megascops kennicottii Megascops asio Megascops seductus Megascops guatemalae Megascops centralis

20. [pp. 323–324] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Moyle 2006) have shown that our current linear sequence of species in the genus *Chloroceryle* does not reflect their evolutionary relationships. These findings result in the following changes:

After the heading and citation for *Chloroceryle*, insert the following:

Notes.—Linear sequence of species follows Moyle (2006).

Rearrange the sequence of species in the genus *Chloroceryle* to:

Chloroceryle amazona Chloroceryle aenea Chloroceryle americana Chloroceryle inda

21. [pp. 236–238] Phylogenetic analyses of mitochondrial DNA sequences (Johansson et al. 2018) have shown that our current linear sequence of species in the genus *Ara* does not reflect their evolutionary relationships. These findings result in the following changes:

After the heading and citation for *Ara*, insert the following:

Notes.—Linear sequence of species follows Johansson et al. (2018).

Rearrange the sequence of species in the genus *Ara* to:

Ara ararauna Ara severus Ara tricolor Ara macao Ara chloropterus Ara militaris Ara ambiguus

22. [p. 239] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Smith et al. 2013) have shown that our current linear sequence of species in the genus *Forpus* does not reflect their evolutionary relationships. These findings result in the following changes:

After the heading and citation for *Forpus*, insert the following:

Notes.—Linear sequence of species follows Smith et al. (2013).

Rearrange the sequence of species in the genus *Forpus* to:

Forpus cyanopygius Forpus passerinus Forpus conspicillatus

23. [p. 240] After the species account for *Brotogeris versicolurus*, insert the following new species account:

Brotogeris chiriri (Vieillot). Yellow-chevroned Parakeet.

Psittacus chiriri Vieillot, 1817 (1818), Nouveau Dictionnaire Histoire Naturelle 25: 359. (Paraguay, *ex* Azara, no. 283.)

Habitat.—Urban and suburban residential areas and parks with diverse exotic tree plantings (palms, *Ceiba*, etc.); in South America, Tropical Deciduous Forest, Gallery Forest, Tropical Lowland Evergreen Forest Edge, Secondary Forest (0–1550 m; Tropical and lower Subtropical zones).

Distribution.—Resident in South America from northern Bolivia and southern Amazonian Brazil south to Paraguay and northern Argentina.

Introduced and established in California (mainly urban coastal slope of Los Angeles County and adjacent western Orange County); introduced populations also present in Miami metropolitan region of Florida, and in the vicinity of Buenos Aires, Argentina.

Notes.—Formerly considered conspecific with *B. versicolurus* (the combined species known as Canarywinged Parakeet), which it has largely replaced in southern California; both species occur in southern Florida, although *chiriri* is increasingly predominant.

Replace the existing Notes for *Brotogeris versicolurus* with the following:

Notes.—See comments under Brotogeris chiriri.

24. [p. 364] Change the English name for *Epinecrophylla fulviventris* to Checker-throated Stipplethroat, following Remsen et al. (2020). Insert the following statement at the beginning of the Notes for this species: Formerly (e.g., AOU 1983, 1998) known as Checker-throated Antwren.

25. [pp. 367–368] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Bravo 2012) have shown that *Myrmeciza* as currently constituted is polyphyletic. These findings result in the following changes:

After the heading and citation for *Myrmeciza*, insert the following:

Notes.—See comments under *Poliocrania*, *Sipia*, and *Myrmeciza zeledoni*.

Insert the following new heading, citation, and Notes after the species account for *Myrmeciza longipes*:

Genus POLIOCRANIA Bravo, Isler, and Brumfield

Poliocrania Bravo, Isler, and Brumfield, 2013, Zootaxa 3717: 488. Type, by original designation, *Myrmeciza exsul* Sclater.

Notes.—The sole species in this genus, *Poliocrania exsul*, was formerly (e.g., AOU 1983, 1998) placed in *Myrmeciza*,

but genetic data (Bravo 2012) indicate that *Myrmeciza* as previously constituted was polyphyletic and that *P. exsul* is not sister to *Myrmeciza sensu stricto*. Separate generic status (Isler et al. 2013) is supported by differences in morphology and behavior from its sister genera *Ampelornis* Isler et al., 2013, and *Sipia*.

Change *Myrmeciza exsul* to *Poliocrania exsul*, add parentheses around the author name in the heading for this species, make the appropriate changes in generic names or abbreviations within the existing Notes, move the account for this species to follow the heading and citation for *Poliocrania*, and add the following sentence to the end of the existing Notes: See comments under *Poliocrania*.

Insert the following new heading, citation, and Notes after the species account for *Poliocrania exsul*:

Genus SIPIA Hellmayr

Sipia Hellmayr, 1924, Field Museum of Natural History Publications, Zoological Series 13, Vol. 3, p. 224. Type, by original designation, *Pyriglena berlepschi* Hartert.

Notes.—Formerly (e.g., AOU 1983, 1998) synonymized with *Myrmeciza*, but genetic data (Bravo 2012) indicate that *Myrmeciza* as previously constituted was polyphyletic and that species placed in *Sipia* are not *Myrmeciza sensu stricto*. Separate generic status (Isler et al. 2013) is supported by differences in morphology and behavior from its sister genus *Ampelornis* Isler et al., 2013.

Change *Myrmeciza laemosticta* to *Sipia laemosticta*, add parentheses around the author name in the heading for this species, move the account for this species to follow the heading and citation for *Sipia*, and add the following sentence to the end of the existing Notes: See comments under *Sipia*.

Move the species account for *Myrmeciza zeledoni* to follow the account for *M. longipes*, and insert the following at the end of the species account for *M. zeledoni*. This species is not related to true *Myrmeciza* and is generally now placed in *Hafferia* (e.g., Remsen et al. 2020), but this generic allocation is being reconsidered by the AOS South American Classification Committee. This species is retained in *Myrmeciza* pending their decision.

26. [p. 370] *Formicarius moniliger* is treated as a species separate from *F. analis*. Change the species account for *F. analis* as follows: delete mention of the *moniliger* group from the distributional statement and change the existing Notes to:

Notes.—Groups: *F. hoffmanni* (Cabanis, 1861) [Hoffmann's Antthrush] and *F. analis* [Black-faced Antthrush]. These groups show differences in plumage and song but the distributional breaks in these characters are not concordant (Howell 1994). See comments under *F. moniliger*.

Insert the following new species account before the account for *E. analis*:

Formicarius moniliger Sclater. Mayan Antthrush.

Formicarius moniliger Sclater, 1856 (1857), Proceedings of the Zoological Society of London 24: 294. (Córdova, Veracruz, Mexico.)

Habitat.—Tropical Lowland Evergreen Forest, Riveredge Forest (0–1850 m; Tropical and Subtropical zones).

Distribution.—*Resident* on the Gulf-Caribbean slope from southern Veracruz, northern Oaxaca, Tabasco, Chiapas, and Yucatan Peninsula south to northern Honduras.

Notes.—Formerly (e.g., AOU 1983, 1998) considered conspecific with *F. analis*, but separated based on differences in song, plumage, and genetics (Howell 1994, Miller 2008, Patten 2015).

27. [p. 352] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Derryberry et al. 2011) have shown that the genus *Philydor* is polyphyletic. These findings result in the following changes:

Replace the existing Notes under Genus *PHILYDOR* Spix with: See comments under *Dendroma*.

Insert the following new heading, citation, and Notes after the species account for *Philydor fuscipenne*:

Genus **DENDROMA** Swainson

Dendroma Swainson, 1837, Natural History and Classification of Birds 2: 316. Type, by monotypy, D. caniceps Swainson = Dendrocopus rufus Vieillot.

Notes.—Formerly (e.g., AOU 1983, 1998) synonymized with *Philydor*, but genetic data (Derryberry et al. 2011) indicate that *Philydor* as previously constituted was polyphyletic and that *Dendroma rufa* is not closely related to *Philydor sensu stricto*.

Change *Philydor rufum* to *Dendroma rufa*, move the account for this species to follow the heading, citation, and

Notes for *Dendroma*, and add the following Notes to the end of the species account:

Notes.—See comments under *Dendroma*.

28. [p. 425] The genus name *Dixiphia* has been determined to be a junior synonym of *Arundinicola* and therefore not available for the species currently listed as *Dixiphia pipra* (Kirwan et al. 2016). This finding results in the following changes:

Delete the heading and citation for *Dixiphia*, and replace the existing Notes under the genus *Ceratopipra* with the Notes currently under *Dixiphia*.

Insert the following new heading, citation, and Notes after the species account for *Manacus vitellinus*:

Genus PSEUDOPIPRA Kirwan et al.

Pseudopipra Kirwan, David, Gregory, Jobling, Steinheimer and Brito, 2016, Zootaxa 4121: 93. Type, by original designation, *Parus pipra*, Linnaeus.

Notes.—The sole species in this genus, *Pseudopipra pipra*, was formerly (e.g., Chesser et al. 2013) placed in *Dixiphia*, but *Dixiphia* is a junior synonym of *Arundinicola* and is therefore not available for this species (Kirwan et al. 2016). This species was also previously placed in *Pipra* (e.g., AOU 1983, 1998). See David et al. (2017) concerning potential use of *Pythis* Boie, 1826. See comments under *Ceratopipra*.

Change *Dixiphia pipra* to *Pseudopipra pipra*, move the account for this species to follow the heading, citation, and Notes for *Pseudopipra*, and replace the last sentence of the Notes with the following: See comments under *Pseudopipra*.

Change the existing Notes under Genus *CERATOPIPRA* Bonaparte, and change the last sentence of the existing Notes for *Ceratopipra mentalis* and *C. erythrocephala* to: See comments under *Pipra*.

29. [p. 380] *Zimmerius parvus* and extralimital species *Z. improbus* and *Z. petersi* are treated as species separate from *Z. vilissimus*. In the account for *Z. vilissimus*, change the English name to Guatemalan Tyrannulet, and replace the existing habitat statement, distributional statement, and Notes with the following:

Habitat.—Montane Evergreen Forest, Tropical Lowland Evergreen Forest Edge, Secondary Forest (500–2600 m; Tropical to lower Temperate zones).

Distribution.—*Resident* in the highlands of eastern Chiapas, Guatemala, and central El Salvador (Sierra de Balsamo).

Notes.—Formerly placed in the genus *Tyranniscus* Cabanis and Heine, 1859. Formerly (e.g., AOU 1983, AOU 1998) considered conspecific with Z. parvus and the extralimital species Z. improbus (Sclater and Salvin, 1871) [Spectacled Tyrannulet] and Z. petersi (Berlepsch, 1907) [Venezuelan Tyrannulet] under the English name Paltry Tyrannulet, but separated from Z. parvus based on differences in plumage, morphometrics, vocalizations, and genetics (Traylor 1982, Rheindt et al. 2013, del Hoyo et al. 2020, Fitzpatrick et al. 2020) and from improbus and petersi based on genetic data (Rheindt et al. 2013) that indicate that these species are not closely related to vilissimus. Details of the distributions of Z. vilissimus and Z. parvus at lower elevations in Guatemala, Belize, and northern Honduras require further study.

Insert the following new species account after the account for *Z. vilissimus*:

Zimmerius parvus (Lawrence). Mistletoe Tyrannulet.

Tyranniscus parvus Lawrence, 1862, Ibis 1862: 12. (Isthmus of Panama; the 2 cotypes are presumably from Lion Hill, Canal Zone.)

Habitat.—Montane Evergreen Forest, Tropical Lowland Evergreen Forest Edge, Secondary Forest (0–3000 m; Tropical to lower Temperate zones).

Distribution.—Resident in the lowlands of southern Belize, eastern Guatemala (southeastern Petén, Izabal), eastern Honduras, and Nicaragua (except Pacific slope), throughout Costa Rica (except dry northwest), Panama, and northwestern Colombia.

Notes.—See comments under *Z. vilissimus*.

30. [p. 428] After the species account for *Lanius cristatus*, insert the following new species account:

Lanius collurio Linnaeus. Red-backed Shrike.

Lanius collurio Linnaeus, 1758, Systema Naturae (ed. 10): 94 (in Europa = Sweden.)

Habitat.—Dry country with low scattered or open growth of thick bushes, shrubs, or low trees, including steppe and scrub desert areas; in Central Asia to subalpine meadows in the Caucasus. In winter, arid savannas with preference for *Acacia*.

Distribution.—*Breeds* nearly throughout Europe (much less common in northwestern Europe) and in Asia east to central Siberia to upper basin of the River Ob and central Altai, to northwest China, and south to some Mediterranean islands (scarce), locally in mountainous

areas of Syria, Lebanon, and Israel, western Turkey, and northeastern Iran to west side of Caspian Sea.

Winters from East Africa at about the equator south through South Africa.

Migrates through northern Africa from the Nile Valley (spring migration more easterly) and east through the Arabian Peninsula, in fall through Afghanistan, northwestern India, and Pakistan.

Casual to northwestern Africa, Canaries, Madeira, Azores, Faeroes, and east to Korea and Japan.

Accidental to Iceland, Madagascar, Marion Island, and Hong Kong.

Accidental in Alaska (Gambell, St. Lawrence Island, Alaska, 3–22 October 2017; photos; Pyle et al. 2018, Lehman et al. 2019).

Notes.—Hybridizes regularly with *L. phoenicuroides* (Schalow, 1875) [Turkestan Shrike] and *L. isabellinus* Hemprich & Ehrenberg, 1833 [Isabelline Shrike] and rarely with Brown Shrike (Worfolk 2000). *Lanius phoenicuroides* and *L. isabellinus* were treated as subspecies of *L. collurio* by Vaurie (1959). A wintering bird from coastal Mendocino County, California, from 5 March–22 April 2015 was determined to be a likely *L. collurio* × *L. phoenicuroides* hybrid (Pyle et al. 2015).

Delete the last sentence of the existing Notes under *Lanius cristatus*.

31. [p. 445] In the original species name *Cyanocorax nanus*, *nanus* is a noun and its ending is not variable (Dickinson and Christidis 2014). Change *Cyanolyca nana* to *Cyanolyca nanus* and insert the following Notes at the end of the species account: Formerly (e.g., AOU 1983, 1998) known as *Cyanolyca nana*, but *nanus* is a noun and not variable (Dickinson and Christidis 2014).

32. [pp. 449–450] *Corvus caurinus* is recognized as representing a geographical trend, rather than a species or subspecies, and thus is treated as a junior synonym of *Corvus brachyrhynchos*, following Slager et al. (2020). Remove the species account for *C. caurinus* and replace the existing habitat statement, distributional statement, and Notes in the account for *C. brachyrhynchos* with the following:

Habitat.—Open forest and woodland used for nesting and roosting, increasing in urban and suburban areas; open and partly open country for foraging, including agricultural lands, urban areas, orchards, and tidal flats; coastal tidelands near coniferous woodland or forest edge in coastal Alaska and Pacific Northwest; restricted mostly to riparian woodland and adjacent areas in arid regions.

Distribution.—*Breeds* along the Pacific coast from south-coastal and southeastern Alaska (west to Kodiak

Island) south through western British Columbia (including Haida Gwaii and Vancouver Island), and from north-central British Columbia, southwestern Northwest Territories, northern Saskatchewan, northern Manitoba, northern Ontario, south-central Quebec, and Newfoundland south to northwestern Baja California (to Ensenada), central Arizona, southern New Mexico, central and southeastern Texas (to Odessa, San Antonio, and north of Corpus Christi), the Gulf coast, and southern Florida (except the Florida Keys).

Winters along the Pacific coast from south-coastal and southeastern Alaska (west to Kodiak Island) south through western British Columbia (including Haida Gwaii and Vancouver Island), and from southern Canada (British Columbia east to Newfoundland) south through the breeding range occasionally to the Florida Keys, and casually to southern Arizona.

Introduced and established on Bermuda.

Casual in southern Nunavut, northwestern Sonora, and western Chihuahua.

Notes.—Formerly (e.g., AOU 1983, 1998) treated as 2 species *C. brachyrhynchos* and *C. caurinus* Baird, 1858 [Northwestern Crow], but merged based on genomic data that indicate a lack of reproductive isolation (Slager et al. 2020), clinal variation, and a lack of consistent differences in size, ecology, and vocalizations (Rhoads 1893, Johnston 1961, Slager et al. 2020). Also known as Common Crow.

33. [pp. 454–457] Phylogenetic analyses of mitochondrial DNA sequences (Sheldon et al. 2005, Moyle et al. 2008) have shown that our current linear sequence of species in the genus *Progne* does not reflect their evolutionary relationships. These findings result in the following changes:

After the heading and citation for *Progne*, insert the following:

Notes.—Linear sequence of species follows Sheldon et al. (2005) and Moyle et al. (2008).

Rearrange the sequence of species in the genus *Progne* to:

Progne tapera
Progne subis
Progne elegans
Progne chalybea
Progne sinaloae
Progne cryptoleuca
Progne dominicensis

34. [p. 515] *Zosterops japonicus* is treated as separate from extralimital species *Z. simplex*. Change the English name of *Z. japonicus* to Warbling White-eye and change the first paragraph of the distributional statement in the account for *Z. japonicus* as follows: "*Resident* from Japan and coastal southern Korean Peninsula, south through the

Ryukyu and Volcano islands, throughout the Philippines, and from central Sumatra through Java, Bali, Sulawesi, the Lesser Sundas, and the southern Moluccas." Replace the existing Notes with the following:

Notes.—Formerly (e.g., AOU 1983, 1998) considered conspecific with *Z. simplex* Swinhoe, 1861 [Swinhoe's White-eye] and known as Japanese White-eye, but separated based on paraphyly of mitochondrial DNA and differences in morphology and vocalizations (Lim et al. 2019, Van Balen 2020).

35. [p. 489] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Alström et al. 2018) have revealed deep divergences within the genus *Locustella* and shown that the linear sequence of species in this genus does not accurately reflect their evolutionary relationships. These findings result in the following changes:

Insert the following new heading, citation, and Notes after the heading **LOCUSTELLIDAE**: Grasshopper-Warblers:

Genus HELOPSALTES Alström et al.

Helopsaltes Alström, Cibois, Irestedt, Zuccon, Fjeldså,
 Andersen, Moyle, Pasquet, and Olsson, 2018,
 Molecular Phylogenetics and Evolution 127: 374. Type,
 by original designation, Motacilla Certhiola Pallas.

Notes.— Formerly (e.g., AOU 1983, 1998) synonymized with *Locustella*, but genetic data (Alström et al. 2018) indicate deep genetic divergences within this genus and show that species placed in *Helopsaltes* are not *Locustella sensu stricto*.

Change *Locustella ochotensis* to *Helopsaltes ochotensis*, move the account for this species to follow the heading and citation for *Helopsaltes*, and replace the second sentence of the existing Notes with the following: See comments under *Helopsaltes*.

After the heading Family **LOCUSTELLIDAE**: Grasshopper-Warblers, insert the following sentence at the end of the existing Notes: Linear sequence of species follows Alström et al. (2018).

After the heading and citation for *Locustella*, insert the following:

Notes.—See comments under *Helopsaltes*.

Rearrange the sequence of species in the family Locustellidae to:

Helopsaltes ochotensis Locustella lanceolata Locustella fluviatilis **36.**[p. 507] *Turdus eunomus* is treated as a species separate from extralimital species *T. naumanni*. Remove the species account for *T. naumanni* and replace it with the following new account:

Turdus eunomus Temminck. Dusky Thrush.

Turdus eunomus Temminck, 1831, Nouveau Recueil de Planches Coloriées d'Oiseaux, livraison 87, text to plate 514. (Japan.)

Habitat.—Open coniferous and mixed forest, forest edge, taiga, and deciduous scrub; in migration and winter, fields, farmland, open woodland, parks, and gardens.

Distribution.—*Breeds* from northern Siberia east to Kamchatka.

Winters from Japan and the Ryukyu Islands south to southern China and Taiwan, rarely west to Southeast Asia and India.

Casual in Alaska (western Aleutians, St. Lawrence Island, Barrow, Anchorage, Sitka), British Columbia (Langley, Nanaimo, Vancouver), the British Isles, western Europe, and the Commander Islands.

Notes.—Formerly (e.g., AOU 1983, 1998) considered conspecific with *T. naumanni* Temminck, 1831 [Naumann's Thrush] under the English name Dusky Thrush, but separated based on evidence of assortative mating in contact zones (Stepanyan 1983 *in* Murray 2009), as in, e.g., Knox et al. (2008), Clements et al. (2019), and Gill et al. (2020). A report of a vagrant individual of *T. naumanni sensu stricto* in Alaska (Gambell, St. Lawrence Island, 5 June 2015; photos; Lehman 2019) is under consideration by the Alaska Checklist Committee.

37. [p. 681] Records of *Uraeginthus bengalus* in the United States are recognized as belonging to populations that were never established (*contra* Long 1981, AOU 1983, 1998; Lever 1987, Pratt et al. 1987, Pyle and Pyle 2009). Remove this species from the list of species known to occur in the United States, remove the heading and citation for *Uraeginthus*, remove the species account from the Main List, and insert the following new species account in the Appendix, Part 1, preceding the account for *Lagonosticta rubricata*:

Uraeginthus bengalus (Linnaeus). Red-cheeked Cordonbleu.

Fringilla bengalus Linnaeus, 1766, Systema Naturae (ed. 12) 1: 323. Based on "Le Bengali" Brisson, Ornithologie 3: 203, pl. 10, fig. 1. (in Bengala error = Senegal.)

This common cagebird, which breeds naturally in tropical Africa, was introduced and formerly bred on the Hawaiian islands of Oahu and Hawaii, especially the

latter (VanderWerf et al. 2018). The small population on Hawaii was believed to have originated in 1972 following the release of aviary birds (Giffin 2003). This population remained stable into the late 1980s, possibly as a result of continued release of captive birds, but declined rapidly in the 21st century and was last sighted in Hawaii in 2006 (VanderWerf et al. 2018). Although formerly treated as established in Hawaii (e.g., Long 1981, AOU 1983, 1998; Lever 1987, Pratt et al. 1987, Pyle and Pyle 2009), this species in fact failed to become established there (Pyle and Pyle 2017, VanderWerf et al. 2017, 2018).

38. [p. 681] Records of *Estrilda melpoda* in the United States are recognized as belonging to populations that are no longer established. Remove this species from the list of species known to occur in the United States and change the second paragraph of the distributional statement to the following:

Introduced and established on Bermuda (reported 1975, well-established and breeding since 1982), and on Puerto Rico. Introduced and formerly (e.g., Long 1981, AOU 1983, 1998; Lever 1987, Pratt et al. 1987, Pyle and Pyle 2009) considered established in the Hawaiian Islands (Oahu and Maui; first reported in 1965), but populations there are no longer considered to be established (Pyle and Pyle 2017).

39. [p. 681] Records of *Estrilda troglodytes* in the United States are recognized as belonging to populations that were never established (Pratt et al. 1987; *contra* Long 1981, AOU 1983, 1998; Lever 1987, Pyle and Pyle 2009). Remove this species from the list of species known to occur in the United States, and change the second paragraph of the distributional statement to the following:

Introduced and established on Puerto Rico. Introduced in the Hawaiian Islands (Oahu and Hawaii, where last reported in 2009), but populations failed to become established (Pyle and Pyle 2017, VanderWerf et al. 2017, 2018).

40. [p. 683] Records of Lonchura malacca in the United States are recognized as belonging to populations that were never established (contra Restall 1996, Banks et al. 2000). Remove this species from the list of species known to occur in the United States, remove "Hawaiian Islands (Oahu)," from the first sentence of the second paragraph of the distributional statement, and add the following sentence to the end of the second paragraph of the distributional statement: Individuals of *L. malacca sensu stricto* are occasionally reported in the Hawaiian Islands (Oahu, Maui), but previous reports of established populations (e.g., Restall 1996, Banks et al. 2000) were in error (Pyle and Pyle 2017). Records from Florida have been accepted by the Florida Ornithological Society Records Committee (Greenlaw 2016) as natural vagrants from established populations in Cuba, but the American Birding Association Checklist Committee has yet to accept these records.

41. [p. 575] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Burns et al. 2014) have shown that *Tachyphonus* as currently constituted is polyphyletic. These findings result in the following changes:

After the species account for *Eucometis penicillata*, insert the following heading, citation, and Notes:

Genus LORIOTUS Jarocki

Loriotus Jarocki, 1821, Zoologiia czyli Zwiérzętopismo Ogólne Podług Naynowszego Systematu Ułożone 2: 133. Type, by original designation, *Tanagra cristata* Linnaeus.

Notes.—Formerly (e.g., AOU 1983, 1998) synonymized with *Tachyphonus*, but genetic data (Burns et al. 2014) indicate that *Tachyphonus* as previously constituted was polyphyletic, and that the species placed in *Loriotus* are not *Tachyphonus sensu stricto*.

Change *Tachyphonus luctuosus* to *Loriotus luctuosus*, place the account for this species under the heading and Notes for *Loriotus*, and insert the following Notes at the end of the species account:

Notes.—See comments under *Loriotus*.

After the heading and citation for *Tachyphonus*, insert the following:

Notes.—See comments under Loriotus.

- **42.** [p. 694] Delete the account for *Brotogeris chiriri* from the Appendix, Part 1.
- **43.** [p. 695] In the Appendix, Part 1, change *Amazilia brevirostris* to *Chrysuronia brevirostris* and *Amazilia tobaci* to *Saucerottia tobaci*, following Stiles et al. (2017).
- **44.** [p. 695] Insert the following new species account in the Appendix, Part 1, preceding the account for *Ramphastos brevis*:

Alcedo atthis (Linnaeus). Common Kingfisher.

Gracula Atthis, Linnaeus, 1758, Systema Naturae (ed. 10) 1: 109. (Egypt.)

A specimen of this species, which naturally occurs in Eurasia and North Africa, was apparently collected from mangroves near Palo Alto, east of Júcaro and south of Ciego de Ávila, Cuba; this specimen was obtained for a private collection on 20 April 2003 and the record was published in Rodríguez et al. (2005). However, no photos were included in the publication, and the identification cannot be verified independently. Rodríguez et al. (2005) did not believe this to have been an escaped cage bird, and the species is occasionally

found far from its typical range (e.g., Iceland, Madeira, Cocos [Keeling] Island, and the Azores), revealing some capacity for dispersal over open water. Nevertheless, whether this individual represented a natural vagrant is uncertain.

45. [pp. 705 ff.] Make the following changes to the list of French names of North American birds:

Insert the following names in the proper position as indicated by the text of this supplement:

Sarkidiornis sylvicola Canard sylvicole Anas diazi Canard du Mexique Selasphorus heloisa Colibri héloïse Selasphorus ellioti Colibri d'Elliot Phaeoptila sordida Colibri sombre Riccordia ricordii Émeraude de Ricord Riccordia bracei Émeraude de New Providence Riccordia swainsonii Émeraude d'Hispaniola Riccordia maugaeus Émeraude de Porto Rico Riccordia bicolor Colibri à tête bleue Cynanthus auriceps Émeraude couronnée Cynanthus forficatus Émeraude de Cozumel Cynanthus canivetii Émeraude de Canivet Basilinna leucotis Colibri à oreilles blanches Basilinna xantusii Colibri de Xantus Pampa curvipennis Campyloptère pampa Pampa excellens Campyloptère de Wetmore Pampa rufa Campyloptère roux Microchera cupreiceps Colibri à tête cuivrée Microchera chionura Colibri elvire Goldmania bella Colibri du Pirré Eupherusa ridgwayi Dryade du Mexique Leucolia violiceps Ariane à couronne violette Leucolia viridifrons Ariane à front vert Saucerottia cyanocephala Ariane à couronne azur Saucerottia hoffmanni Ariane de Hoffmann Saucerottia beryllina Ariane béryl Saucerottia cvanura Ariane à queue bleue Saucerottia edward Ariane d'Edward Chrysuronia coeruleogularis Colibri faux-saphir Chrysuronia humboldtii Ariane de Humboldt Polyerata amabilis Ariane aimable Polverata decora Ariane charmante Chlorestes candida Ariane candide Chlorestes eliciae Colibri d'Élicia Chlorestes julie Colibri de Julie Buteo rufinus Buse féroce Gymnasio nudipes Petit-duc de Porto Rico Brotogeris chiririri Toui à ailes jaunes Poliocrania exsul Alapi à dos roux Sipia laemosticta Alapi tabac Formicarius moniliger Tétéma du Mexique Dendroma rufa Anabate roux Pseudopipra pipra Manakin à tête blanche

Zimmerius parvus Tyranneau menu
Lanius collurio Pie-grièche écorcheur
Cyanolyca nanus Geai nain
Helopsaltes ochotensis Locustelle de Middendorff
Turdus eunomus Grive à ailes rousses
Loriotus luctuosus Tangara à épaulettes blanches
in APPENDIX (Part 1)
Saucerottia tobaci Ariane de Félicie
Chrysuronia brevirostris Ariane à poitrine blanche
Alcedo atthis Martin-pêcheur d'Europe
Uraeginthus bengalus Cordonbleu à joues rouges

Delete the following names:

Sarkidiornis melanotos Canard à bosse Atthis heloisa Colibri héloïse Atthis ellioti Colibri d'Elliot Chlorostilbon auriceps Émeraude couronnée Chlorostilbon forficatus Émeraude de Cozumel Chlorostilbon canivetii Émeraude de Canivet Chlorostilbon ricordii Émeraude de Ricord Chlorostilbon bracei Émeraude de New Providence Chlorostilbon swainsonii Émeraude d'Hispaniola Chlorostilbon maugaeus Émeraude de Porto Rico Cynanthus sordidus Colibri sombre Cyanophaia bicolor Colibri à tête bleue Campylopterus curvipennis Campyloptère pampa Campylopterus excellens Campyloptère de Wetmore Campylopterus rufus Campyloptère roux *Elvira chionura* Colibri elvire Elvira cupreiceps Colibri à tête cuivrée *Thalurania ridgwayi* Dryade du Mexique Amazilia candida Ariane candide Amazilia amabilis Ariane aimable Amazilia decora Ariane charmante Amazilia cyanocephala Ariane à couronne azur Amazilia beryllina Ariane béryl *Amazilia cyanura* Ariane à queue bleue Amazilia hoffmanni Ariane de Hoffmann Amazilia edward Ariane d'Edward *Amazilia violiceps* Ariane à couronne violette Amazilia viridifrons Ariane à front vert Goethalsia bella Colibri du Pirré Lepidopyga coeruleogularis Colibri faux-saphir *Juliamyia julie* Colibri de Julie Hylocharis humboldtii Saphir de Humboldt Hylocharis eliciae Saphir d'Elicia *Hylocharis leucotis* Saphir à oreilles blanches Hylocharis xantusii Saphir de Xantus Megascops nudipes Petit-duc de Porto Rico Myrmeciza exsul Alapi à dos roux Myrmeciza laemosticta Alapi tabac Philydor rufa Anabate roux Dixiphia pipra Manakin à tête blanche

Cyanolyca nana Geai nain
Corvus caurinus Corneille d'Alaska
Locustella ochotensis Locustelle de Middendorff
Turdus naumanni Grive de Naumann
Uraeginthus bengalus Cordonbleu à joues rouges
Tachyphonus luctuosus Tangara à épaulettes blanches
in APPENDIX (Part 1)
Amazilia brevirostris Ariane à poitrine blanche

Amazilia brevirostris Ariane à poitrine blanche *Amazilia tobaci* Ariane de Félicie *Brotogeris chiririri* Toui à ailes jaunes

Change the sequence of families in the order Suliformes as indicated by the text of this supplement.

Change the sequence of genera and species in the families PHASIANIDAE, TROCHILIDAE, RALLIDAE, PHALACROCORACIDAE, CATHARTIDAE, STRIGIDAE, ALCEDINIDAE, PSITTACIDAE, THAMNOPHILIDAE, HIRUNDINIDAE, and LOCUSTELLIDAE as indicated by the text of this supplement.

Proposals considered but not accepted by the Committee included recognition of the columbid subfamily Starnoenadinae, change of the English name of Blue-headed Quail-Dove Starnoenas cyanocephala, separation of Garnet-throated Hummingbird Lamprolaima rhami into 2 species, recognition of Guanacaste Hummingbird Amazilia alfaroana as a species rather than a hybrid, separation of Ardea occidentalis from Great Blue Heron A. herodias, separation of Aegolius brooksi from Northern Saw-whet Owl A. acadicus, removal of "scrub" from the English names of the scrubjays (Aphelocoma spp.), separation of Unicolored Jay Aphelocoma unicolor into 4 species, separation of Horned Lark Eremophila alpestris into 2 or more species, and change of the English name of Olive Warbler Peucedramus taeniatus to Ocotero.

ACKNOWLEDGMENTS

Normand David serves as the Committee's advisor for classical languages in relation to scientific names, and Michel Gosselin is the authority for French names. Natalia C. García, Rosa A. Jimenez, and Max T. Kirsch serve on the Early Professional Systematics Group for the committee. We thank P. Alström, G. R. Angehr, F. Angulo, N. Bahr, L. Bevier, M. Brown, S. Claramunt, D. DeRaad, E. C. Dickinson, K. L. Garrett, V. Ruiz-Gutiérrez, G. Hanisek, M. J. Iliff, M. L. Isler, A. Jaramillo, A. W. Jones, A. J. Knue, M. Kuziemko, P. E. Lehman, D. Lepage, T. Leukering, A. Lin-Moore, J. Maley, J. McCormack, S. Mlodinow, P. Moriyasu, K. E. Omland, F. Pacheco, A. P. Peterson, P. Pyle, V. de Q. Piacentini, M. L. P. Retter, R. Schodde, T. S. Schulenberg, D. L. Slager, G. Stiles, T. Valqui, B. Winger, J. Withrow, R. Wright, and L. Zoller for assistance, suggestions, and comments.

LITERATURE CITED

- Alström, P., A. Cibois, M. Irestedt, D. Zuccon, M. Gelang, J. Fjeldså, M. J. Andersen, R. G. Moyle, E. Pasquet, and U. Olsson (2018). Comprehensive molecular phylogeny of the grassbirds and allies (Locustellidae) reveals extensive non-monophyly of traditional genera, and a proposal for a new classification. Molecular Phylogenetics and Evolution 127:367–375.
- American Ornithologists' Union (1973). Thirty-second supplement to the American Ornithologists' Union Check-List of North American Birds. The Auk 90:411–419.
- American Ornithologists' Union (1983). Check-list of North American Birds, 6th ed. American Ornithologists' Union, Lawrence, KS, USA.
- American Ornithologists' Union (1998). Check-list of North American Birds, 7th ed. American Ornithologists' Union, Washington, DC, USA.
- Ankney, C. D., and D. G. Dennis (1988). Response to Hepp *et al*. The Auk 105:807–808.
- Ankney, C. D., D. G. Dennis, L. N. Wighard, and J. E. Seeb (1986). Low genic variation between Black Ducks and Mallards. The Auk 103:701–709.
- Avise, J. C., C. D. Ankney, and W. S. Nelson (1990). Mitochondrial gene trees and the evolutionary relationship of Mallard and Black Ducks. Evolution 44:1109–1119.
- Banks, R. C., C. Cicero, J. L. Dunn, A. W. Kratter, H. Ouellet, P. C. Rasmussen, J. V. Remsen, J. A. Rising, and D. F. Stotz Jr. (2000). Forty-second supplement to the American Ornithologists' Union Check-List of North American Birds. The Auk 117:847–856.
- Banks, R.C., C. Cicero, J. L. Dunn, A. W. Kratter, P. C. Rasmussen, J. V. Remsen, Jr., J. A. Rising, and D. F. Stotz (2003). Forty-fourth supplement to the American Ornithologists' Union Check-list of North American Birds. The Auk 120:923–931.
- Bellrose, F. H. (1976). Ducks, Geese and Swans of North America. Stackpole Books, Harrisburg, PA, USA.
- Berger, A. J. (1981). Hawaiian Birdlife, second edition. University of Hawaii Press, Honolulu, HI, USA.
- Bravo, G. A. (2012). Phenotypic and niche evolution in the antbirds (Aves, Thamnophilidae). Ph.D. dissertation, Louisiana State University, Baton Rouge, LA, USA.
- Brodsky, L. M., and P. J. Weatherhead (1984). Behavioral and ecological factors contributing to American Black Duck–Mallard hybridization. The Journal of Wildlife Management 48:846–852.
- Brodsky, L. M., C. D. Ankney, and D. G. Dennis (1988). The influence of male dominance on social interactions in Black Ducks and Mallards. Animal Behavior 36:1371–1378
- Brown, D. E. (1985). Arizona Wetlands and Waterfowl. University of Arizona Press, Tucson, AZ, USA.
- Burns, K. J., A. J. Shultz, P. O. Title, N. A. Mason, F. K. Barker, J. Klicka, S. M. Lanyon, and I. J. Lovette (2014). Phylogenetics and diversification of tanagers (Passeriformes: Thraupidae), the largest radiation of Neotropical songbirds. Molecular Phylogenetics and Evolution 75:41–77.
- Chesser, R. T., R. C. Banks, F. K. Barker, C. Cicero, J. L. Dunn, A. W. Kratter, I. J. Lovette, P. C. Rasmussen, J. V. Remsen, Jr., J. D. Rising, D. F. Stotz, and K. Winker (2013). Fifty-fourth

- supplement to the American Ornithologists' Union Check-list of North American Birds. The Auk 130:558–571.
- Chesser, R. T., K. J. Burns, C. Cicero, J. L. Dunn, A. W. Kratter, I. J. Lovette, P. C. Rasmussen, J. V. Remsen, Jr., J. D. Rising, D. F. Stotz, and K. Winker (2017). Fifty-eighth supplement to the American Ornithological Society's Check-list of North American Birds. The Auk: Ornithological Advances 134:751–773.
- Clements, J. F., T. S. Schulenberg, M. J. Iliff, S. M. Billerman, T. A. Fredericks, B. L. Sullivan, and C. L. Wood (2019). The eBird/Clements Checklist of Birds of the World: v2019. https://www.birds.cornell.edu/clementschecklist/download/
- Collinson, J. M., P. Dufour, A. A. Hamza, Y. Lawrie, M. Elliott, C. Barlow, and P-A. Crochet (2017). When morphology is not reflected by molecular phylogeny: The case of three 'orange-billed terns' *Thalasseus maximus, Thalasseus bergii*, and *Thalasseus bengalensis* (Charadriiformes: Laridae). Biological Journal of the Linnean Society 121:439–445.
- Crowe, T., R. Bowie, P. Bloomer, T. Mandiwana, T. Hedderson, E. Randi, S. Pereira, and J. Wakeling (2006). Phylogenetics, biogeography and classification of, and character evolution in, gamebirds (Aves: Galliformes): Effects of character exclusion, data partitioning and missing data. Cladistics 22:495–532.
- Dantas, S. M., J. D. Weckstein, J. M. Bates, N. K. Krabbe, C. D. Cadena, M. B. Robbins, E. Valderrama, and A. Aleixo (2016). Molecular systematics of the New World screech-owls (*Megascops*: Aves, Strigidae): Biogeographic and taxonomic implications. Molecular Phylogenetics and Evolution 94:626–634.
- David, N., S. M. S. Gregory, G. M. Kirwan, J. A. Jobling, F. D. Steinheimer, and G. R. R. Brito (2017). Addendum to Kirwan et al. (2016, Zootaxa 4121(1): 89–94). Zootaxa 4216:299–300.
- Delacour, J. T., and E. Mayr (1945). The family Anatidae. The Wilson Bulletin 57:3–55.
- del Hoyo, J., N. Collar, and G. M. Kirwan (2020). Mistletoe Tyrannulet (*Zimmerius parvus*). In Handbook of the Birds of the World Alive (J. del Hoyo, A. Elliott, J. Sargatal, D. A. Christie, and E. de Juana, Editors). Lynx Edicions, Barcelona, Spain. https://www.hbw.com/species/mistletoe-tyrannulet-zimmerius-parvus
- Derryberry, E. P., S. Claramunt, G. Derryberry, R. T. Chesser, J. Cracraft, A. Aleixo, J. Pérez-Emán, J. V. Remsen, Jr., and R. T. Brumfield (2011). Lineage diversification and morphological evolution in a large-scale continental radiation: The Neotropical ovenbirds and woodcreepers (Aves: Furnariidae). Evolution 65:2973–2986.
- Dickinson, E. C., and L. Christidis (Editors) (2014). The Howard and Moore Complete Checklist of the Birds of the World, 4th edition: Vol. 2, Passerines. Aves Press, Eastbourne, UK.
- Dufour, P., and P.-A. Crochet (2020). Identification of American Royal Tern and African Royal Tern based on photographs and sound-recordings. Dutch Birding 42:1–24.
- Ericson, P. G. P., C. L. Anderson, T. Britton, A. Elzanowski,
 U. S. Johansson, M. Källersjö, J. I. Ohlson, T. J. Parsons,
 D. Zuccon, and G. Mayr (2006). Diversification of Neoaves:
 Integration of molecular sequence data and fossils. Biology
 Letters 2:543–547.
- Fitzpatrick, J., D. A. Christie, and G. M. Kirwan (2020). Paltry Tyrannulet (*Zimmerius vilissimus*). In Handbook of the Birds of the World Alive (J. del Hoyo, A. Elliott, J. Sargatal, D. A. Christie, and E. de Juana, Editors). Lynx Edicions,

- Barcelona, Spain. https://www.hbw.com/species/paltry-tyrannulet-zimmerius-vilissimus
- Ford, R. J., W. Selman, and S. S. Taylor (2017). Hybridization between Mottled Ducks (*Anas fulvigula maculosa*) and Mallards (*A. platyrhynchos*) in the western Gulf Coast region. The Condor: Ornithological Applications 119:683–696.
- Garcia-R, J. C., G. C. Gibb, and S. A. Trewick (2014). Deep global evolutionary radiation in birds: Diversification and trait evolution in the cosmopolitan bird family Rallidae. Molecular Phylogenetics and Evolution 81:96–108.
- Garcia-R, J. C., E. M. Lemmon, A. R. Lemmon, and N. French (2020). Phylogenomic reconstruction sheds light on new relationships and timescale of rails (Aves: Rallidae) evolution. Diversity 12: 70.
- Gibson, D. D., L. H. DeCicco, R. E. Gill, S. C. Heinl, A. J. Lang, T. G. Tobish, Jr., and J. J. Withrow (2013). Third report of the Alaska Checklist Committee, 2008–2012. Western Birds 44:183–195.
- Giffin, J. G. (2003). Pu'u Wa'awa'a Biological Assessment. State of Hawaii, Department of Land and Natural Resources, Division of Forestry and Wildlife, Honolulu, HI, USA.
- Gill F. B., D. Donsker, and P. C. Rasmussen (Editors) (2020). IOC World Bird List (v10.1). doi:10.14344/IOC.ML.10.1
- Greenlaw, J. (2016). Twenty-fourth Report of the Florida Ornithological Society Records Committee: 2016. Florida Field Naturalist 44:29–44.
- Hackett, S. J., R. T. Kimball, S. Reddy, R. C. K. Bowie, E. L. Braun, M. J. Braun, J. L. Chojnowski, W. A. Cox, K.-L. Han, J. Harshman, et al. (2008). A phylogenomic study of birds reveals their evolutionary history. Science 320:1763–1768.
- Hepp, G. R., J. M. Novak, K. T. Scribner, and P. W. Swangel (1988). Genetic distance and hybridization of Black Ducks and Mallards: A morph of a different color? The Auk 105:804–807.
- Hernández-Baños, B. E., L. E. Zamudio-Beltran, and B. Milá (2020). Phylogenetic relationships and systematics of a subclade of Mesoamerican emerald hummingbirds (Aves: Trochilidae: Trochilini). Zootaxa 4748:581–591.
- Hosner, P. A., B. C. Faircloth, T. C. Glenn, E. L. Braun, and R. T. Kimball (2016). Avoiding missing data biases in phylogenomic inference: An empirical study in the landfowl (Aves: Galliformes). Molecular Biology and Evolution 33:1110–1125.
- Howell, S. N. G. (1994). The specific status of Black-faced Antthrushes in Middle America. Cotinga 1:21–25.
- Howell, S. N. G., and S. Webb (1995). A Guide to the Birds of Mexico and Northern Central America. Oxford University Press, New York, NY, USA.
- Hubbard, J. P. (1977). The biological and taxonomic status of the Mexican Duck. Bulletin of the New Mexico Department of Game and Fish 16, Albuquerque, NM, USA.
- Isler, M. L., G. A. Bravo, and R. T. Brumfield (2013). Taxonomic revision of *Myrmeciza* (Aves: Passeriformes: Thamnophilidae) into 12 genera based on phylogenetic, morphological, behavioral, and ecological data. Zootaxa 3717:469–497.
- Johansson, U. S., P. G. P. Ericson, M. P. K. Blom, and M. Irestedt (2018). The phylogenetic position of the extinct Cuban Macaw *Ara tricolor* based on complete mitochondrial genome sequences. Ibis 160:666–672.
- Johnsgard, P. A. (1961). The taxonomy of the Anatidae—A behavioural analysis. Ibis 103A:71–85.

- Johnsgard P. A. (1967). Sympatry changes and hybridization incidence in Mallards and Black Ducks. American Midland Naturalist 77:51–63.
- Johnson, J. A., J. W. Brown, J. Fuchs, and D. P. Mindell (2016). Multilocus phylogenetic inference among New World Vultures (Aves: Cathartidae). Molecular Phylogenetics and Evolution 105:193–199.
- Johnston, D. W. (1961). The Biosystematics of American Crows. University of Washington Press, Seattle, WA, USA.
- Kaplan, J., and G. Hanisek (2012). Seventeenth report of the Avian Records Committee of Connecticut. Connecticut Warbler 32:33–50.
- Kennedy, M., and H. G. Spencer (2014). Classification of the cormorants of the world. Molecular Phylogenetics and Evolution 79:249–257.
- Kirby, R. E., G. A. Sargeant, and D. Shutler (2004). Haldane's rule and American Black Duck × Mallard hybridization. Canadian Journal of Zoology 82:1827–1831.
- Kirwan, G. M., N. David, S. M. S. Gregory, J. A. Jobling, F. D. Steinheimer, and G. R. R. Brito (2016). The mistaken manakin: A new genus-group name for *Parus pipra* Linnaeus, 1758 (Aves: Passeriformes: Pipridae). Zootaxa 4142:89–94.
- Knox, A. G., J. M. Collinson, D. T. Parkins, G. Sangster, and L. Svensson (2008). Taxonomic recommendations for British Birds. Fifth report. Ibis 150:833–835.
- Kratter, A. W., M. Gomes, and K. Matera (2019). First Florida record of Dark-billed Cuckoo (Coccyzus melacorhyphus). Florida Field Naturalist 48:55–59.
- Lavretsky, P., J. M. DaCosta, B. E. Hernández-Baños, A. Engilis, Jr., M. D. Sorenson, and J. L. Peters (2015). Speciation genomics and a role for the Z chromosome in the early stages of divergence between Mexican Ducks and Mallards. Molecular Ecology 24:5364–5378.
- Lavretsky, P., J. M. DaCosta, M. D. Sorenson, K. G. McCracken, and J. L. Peters (2019a). ddRAD-seq data reveal significant genome-wide population structure and divergent genomic regions that distinguish the Mallard and close relatives in North America. Molecular Ecology 28:2594–2609.
- Lavretsky, P., B. E. Hernández-Baños, and J. L. Peters (2014a). Rapid radiation and hybridization contribute to weak differentiation and hinder phylogenetic inferences in the New World Mallard complex (*Anas* spp.). The Auk: Ornithological Advances 131:524–538.
- Lavretsky, P., T. Janzen, and K. G. McCracken (2019b). Identifying hybrids and the genomics of hybridization: Mallards and American Black Ducks of eastern North America. Ecology and Evolution 9:3470–3490.
- Lavretsky, P., K. G. McCracken, and J. L. Peters (2014b). Phylogenetics of a recent radiation in the Mallard and allies (Aves: Anas): Inferences from a genomic transect and the multispecies coalescent. Molecular Phylogenetics and Evolution 70:402–411.
- Lehman, P. (2019). The Birds of Gambell and St. Lawrence Island, Alaska. Studies of Western Birds 4. Western Field Ornithologists, Camarillo, CA, USA.
- Lehman, P., P. Pyle, N. Moores, J. Hough, and G. H. Rosenberg (2019). First North America record of Red-backed Shrike (*Lanius collurio*). North American Birds 70:252–262.
- Lever, C. (1987). Naturalized Birds of the World. Longman Scientific and Technical, Harlow, UK.

- Licona-Vera, Y., and J. F. Ornelas (2017). The conquering of North America: Dated phylogenetic and biogeographic inference of migratory behavior in bee hummingbirds. BMC Evolutionary Biology 17:126.
- Lim, B. T. M., K. R. Sadanandan, C. Dingle, Y. Y. Leung, D. M. Prawiradilaga, M. Irham, H. Ashari, J. G. H. Lee, and F. E. Rheindt (2019). Molecular evidence suggests radical revisions of species limits in the great speciator white-eye genus *Zosterops*. Journal of Ornithology 160:1–16.
- Lockwood, M. W. (1999). Texas Bird Records Committee report for 1998. Bulletin of the Texas Ornithological Society 32:26–37.
- Long, J. L. (1981). Introduced Birds of the World. Universe Books, New York, NY, USA.
- Mactavish, B. (1994). Eurasian Oystercatcher, first for North America. Birders Journal 3:168–171.
- McCracken, K. G., W. P. Johnson, and F. H. Sheldon (2001). Molecular population genetics, phylogeography, and conservation biology of the Mottled Duck (*Anas fulvigula*). Conservation Genetics 2:87–102.
- McGuire, J. A., C. C. Witt, J. V. Remsen, Jr., A. Corl, D. L. Rabosky, D. L. Altshuler, and R. Dudley (2014). Molecular phylogenetics and the diversification of hummingbirds. Current Biology 24:1–7.
- Miller, M. J. (2008). Evolutionary ecological genetics of some Neotropical birds. Ph.D. dissertation, University of Alaska Fairbanks, Fairbanks, AK, USA.
- Moyle, R. G. (2006). A molecular phylogeny of kingfishers (Alcedinidae) with insights into early biogeographic history. The Auk 123:487–499.
- Moyle, R. G., B. Slikas, L. A. Whittingham, D. W. Winkler, and F. H. Sheldon (2008). DNA sequence assessment of phylogenetic relationships among New World martins (Hirundinidae: *Progne*). The Wilson Journal of Ornithology 120:683–691.
- Murray, K. (2009). Naumann's Thrush in Essex: New to Britain. British Birds 102:435–440.
- Özdikmen, H. (2008). *Neodamophila* nom. nov., a replacement name for the bird genus *Damophila* Reichenbach, 1854 (Aves: Apodiformes: Trochilidae). Munis Entomology and Zoology 3:171–173.
- Patten, M. A. (2015). Black-faced Antthrush (Formicarius analis), version 1.0. In Neotropical Birds Online (T. S. Schulenberg, editor). Cornell Lab of Ornithology, Ithaca, NY, USA. https://doi. org/10.2173/nb.blfant1.01
- Piston, A. W., and S. C. Heinl (2001). First record of the European Golden-Plover (*Pluvialis apricaria*) from the Pacific. Western Birds 32:179–181.
- Pranty, B., J. Dunn, S. C. Heinl, A. W. Kratter, P. E. Lehman, M. W. Lockwood, B. Mactavish, and K. J. Zimmer (2008). ABA Checklist: Birds of the Continental United States and Canada, 7th edition. American Birding Association, Colorado Springs, CO, USA.
- Pratt, H. D., P. Bruner, and D. G. Berrett (1987). The Birds of Hawaii and the Tropical Pacific. Princeton University Press, Princeton, NJ, USA.
- Proudfoot, G. A., F. R. Gehlbargh, and R. L. Honeycutt (2007). Mitochondrial DNA variation and phylogeography of the eastern and western screech-owls. The Condor 109: 617–627.

- Prum, R. O., J. S. Berv, A. Dornburg, D. J. Field, J. P. Townsend, E. M. Lemmon, and A. R. Lemmon (2015). A comprehensive phylogeny of birds (Aves) using targeted next-generation DNA sequencing. Nature 526:569–573.
- Pyle, P., M. Gustafson, T. Johnson, A. W. Kratter, A. Lang, K. Nelson, M. W. Lockwood, and D. Sibley (2018). 29th report of the ABA Checklist Committee 2018. Birding 50:30–40.
- Pyle, P., M. Gustafson, T. Johnson, A. W. Kratter, A. Lang, K. Nelson, M. W. Lockwood, and D. Sibley (2019). 30th report of the ABA Checklist Committee, 2019. Birding 51:36–42.
- Pyle, P., R. J. Keiffer, J. L. Dunn, and N. Moores (2015). The Mendocino shrike: Red-backed Shrike (*Lanius collurio*) × Turkestan Shrike (*L. phoenicuroides*) hybrid. North American Birds 69:4–35.
- Pyle, R. L., and P. Pyle (2009). The Birds of the Hawaiian Islands: Occurrence, History, Distribution, and Status, version 1. B. P. Bishop Museum, Honolulu, HI, USA.
- Pyle, R. L., and P. Pyle (2017). The Birds of the Hawaiian Islands: Occurrence, History, Distribution, and Status, version 2. B. P. Bishop Museum, Honolulu, HI, USA. http://hbs.bishopmuseum.org/birds/rlp-monograph
- Remsen, J. V., Jr., J. I. Areta, E. Bonaccorso, S. Claramunt, A. Jaramillo, J. F. Pacheco, M. B. Robbins, F. G. Stiles, D. F. Stotz, and K. J. Zimmer (2020). A classification of the bird species of South America, version 1. American Ornithological Society. http://www.museum.lsu.edu/~Remsen/SACCBaseline.htm
- Restall, R. (1996). Munias and Mannikins. Yale University Press, New Haven, CT, USA.
- Rheindt, F. E., A. M Cuervo, and R. T. Brumfield (2013). Rampant polyphyly indicates cryptic diversity in a clade of Neotropical flycatchers (Aves: Tyrannidae). Biological Journal of the Linnean Society 108:889–900.
- Rhoads, S. N. (1893). Notes on certain Washington and British Columbia birds. The Auk 10:16–24.
- Roberson, D. (1993). Fourteenth report of the California Bird Records Committee. Western Birds 24:113–166.
- Rodríguez, Y., O. H. Garrido, J. W. Wiley, and A. Kirkconnell (2005). The Common Kingfisher (*Alcedo atthis*): An exceptional first record for the West Indies and the Western Hemisphere. Ornitologia Neotropical 16:41.
- Salter, J. F., C. H. Oliveros, P. A. Hosner, J. D. Manthey, M. B. Robbins, R. G. Moyle, R. T. Brumfield, and B. C. Faircloth (2020). Extensive paraphyly in the typical owl family (Strigidae). The Auk: Ornithological Advances 137:1–15.
- Sheldon, F. H., L. A. Whittingham, R. G. Moyle, B. Slikas, and D.W.Winkler (2005). Phylogeny of swallows (Aves: Hirundinidae) estimated from nuclear and mitochondrial DNA sequences. Molecular Phylogenetics and Evolution 35:254–270.
- Slager, D. L., K. L. Epperly, R. R. Ha, S. Rohwer, C. Van Hemert, and J. Klicka (2020). Cryptic and extensive hybridization between ancient lineages of American Crows. Molecular Ecology 29:956–969.
- Smith, B. T., C. C. Ribas, B. M. Whitney, B. E. Hernández-Baños, and J. Klicka (2013). Identifying biases at different spatial and temporal scales of diversification: A case study in the Neotropical parrotlet genus *Forpus*. Molecular Ecology 22:483–494.
- Stepanyan, L. S. (1983). [Superspecies and sibling species in the avifauna of the USSR]. In Russian. Nauka Press, Moscow, Russia.

- Stiles, F. G., J. V. Remsen, Jr., and J. A. McGuire (2017). The generic classification of the Trochilini (Aves: Trochilidae): Reconciling classification with phylogeny. Zootaxa 4353:401–424.
- Traylor, M. A., Jr. (1982). Notes on tyrant flycatchers (Aves: Tyrannidae). Fieldiana (Zoology), New Series 13:1–22.
- Tsai, W. L. E., C. Mota-Vargas, O. Rojas-Soto, R. Bhowmik, E. Y. Liang, J. M. Maley, E. Zarza, and J. E. McCormack (2019). Museum genomics reveals the speciation history of *Dendrortyx* wood-partridges in the Mesoamerican highlands. Molecular Phylogenetics and Evolution 136:29–34.
- Van Balen, B. (2020). Japanese White-eye (*Zosterops japonicus*). In Handbook of the Birds of the World Alive (J. del Hoyo, A. Elliott, J. Sargatal, D. A. Christie, and E. de Juana, Editors). Lynx Edicions, Barcelona, Spain. https://www.hbw.com/species/japanese-white-eye-zosterops-japonicus
- VanderWerf, E. A., R. E. David, P. Donaldson, R. May, H. D. Pratt, P. Pyle, and L. Tanino (2017). Hawaiian Islands bird checklist 2017. 'Elepaio 77:33–42.

- VanderWerf, E. A., R. E. David, P. Donaldson, R. May, H. D. Pratt, P. Pyle, and L. Tanino (2018). First report of the Hawaii Bird Records Committee. Western Birds 49:2–23.
- Vaurie, C. (1959). The Birds of the Palearctic Fauna. Passeriformes. H. F. and G. Witherby, London, UK.
- Walker, R. L. (1967). A brief history of exotic game bird and mammal introductions into Hawaii with a look to the future. 'Elepaio 28:29–32, 39–43.
- Wang, N., R. T. Kimball, E. L. Braun, B. Liang, and Z. Zhang (2013). Assessing phylogenetic relationships among Galliformes: A multigene phylogeny with expanded taxon sampling in Phasianidae. PLOS One 8:e64312.
- Wink, M., A. El-Sayed, H. Sauer-Gürth, and J. Gonzalez (2009). Molecular phylogeny of owls (Strigiformes) inferred from DNA sequences of the mitochondrial cytochrome b and the nuclear RAG-1 gene. Ardea 97:581–590.
- Worfolk, T. (2000). Identification of Red-backed, Isabelline and Brown shrikes. Dutch Birding 22:323–362.